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(54) Title: POLYPEPTIDE HAVING AN ACTIVITY TO SUPPORT PROLIFERATION OR SURVIVAL OF HEMATOPOIETIC STEM CELL AND HEMATOPOIETIC PROGENITOR CELL, AND DNA CODING FOR THE SAME

(57) Abstract: A gene encoding a polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells is isolated by comparing expressed genes between cells which support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells and cells which do not support the proliferation or survival. Proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells is supported by using stromal cells in which the isolated gene is expressed or a gene product of the isolated gene.

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DESCRIPTION

POLYPEPTIDE HAVING AN ACTIVITY TO SUPPORT PROLIFERATION
OR SURVIVAL OF HEMATOPOIETIC STEM CELL AND HEMATOPOIETIC
PROGENITOR CELL, AND DNA CODING FOR THE SAME

Background of the Invention

Field of the Invention

The present invention relates to a polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, a DNA coding the polypeptide, and a pharmaceutical composition comprising the polypeptide as active ingredient.

Description of the Related Art

Fully differentiated mature hematopoietic cells 15 have limited short lives. Homeostasis of the blood is maintained due to supply of the mature blood cells caused by continuous differentiation of hematopoietic progenitor cells. The hematopoietic progenitor cells are giving rise from more undifferentiated 20 hematopoietic stem cells. The hematopoietic stem cells have potential of differentiating into all of the differentiation lineages (totipotency) and have potential of self-renew with retaining the totipotency so as to supply the hematopoietic cells through life. 25 That is, the hematopoietic stem cells are known to generate totipotent stem cells by the self-renew and to



differentiate in parts to a variety of the mature blood cells through the hematopoietic progenitor cells.

This differentiation of the blood cells is regulated by a variety of cytokines. Erythropoietin is known to 5 promote the differentiation of the erythrocytic lineages. G-CSF and thrombopoietin are also known to promote the differentiation of the neutrophils, and the megakaryocytes and the platelet productive cells, respectively. However, a factor required for the selfrenew of the hematopoietic stem cell with retaining the 10 totipotency has not been clear. Although SCF/MGF (Williams, D.E., Cell, 63: 167-174, 1990; Zsebo, K.M., Cell, 63: 213-224, 1990), SCGF (WO98/08869), and the like are reported as growth factors for the hematopoietic stem cells, none of them have potency to 15 sufficiently retain the totipotency of the hematopoietic stem cells. Although attempts to culture the hematopoietic stem cells in the presence of combinations of known cytokines, a system for efficient amplification of the hematopoietic stem cells was not realized (Miller, C. L., Proc. Natl. Acad. Sci. USA, 94: 13648-13653, 1997; Yagi, M., Proc. Natl. Acad. Sci. USA, 96: 8126-8131, 1999; Shih, C.C., Blood, 94: 5 1623-1636, 1999).

On the other hand, attempts to allow the

25 hematopoietic stem cells to survive or proliferate

without differentiation by using stromal cells which

supply an environment suitable for survival or

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proliferation of the hematopoietic stem cells were reported (Moore K.A., Blood, 89: 12, 4337-4347, 1997). In addition, WO99/03980 discloses a stromal cell line capable of supporting proliferation or survival of hematopoietic stem cells and hematopoietic progenitor cells, which are established from an AGM (Aorta-Gonad-Mesonephros) region of a fetal mouse.

It is postulated that there should be more peptides that efficiently facilitate hematopoietic stem cell and progenitor cell amplification by themselves or in combination with stromal cells or stimulating factors such as cytokines, in addition to known factors affecting hematopoietic cells.

Summary of the Invention

Since the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells in vitro can be supported by co-culture of stromal cells and hematopoietic stem cells and hematopoietic progenitor cells, the stromal cells are expected to produce factors supporting the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells. An object of the present invention is to provide a factor supporting the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, which is derived from the stromal cells.

The inventor of the present invention has assumed

that the mouse stromal cells produce factors supporting the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, as mentioned above. Attention is given that there are two kinds of stromal cells. One has a ability to support the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells (hereafter sometimes referred to as "activity to support hematopoietic stem cells"). The other does not have the activity to 10 support hematopoietic stem cells. The inventor of the present invention has assumed that this difference in the ability is due to the fact that expression of genes encoding the factors is increased in the supporting stromal cells and that the expression is low in nonsupporting stromal cells. Thus the inventor think it can be found the factors supporting the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells among the genes expressed higher in the supporting cells compared to in the non-supporting cells. In this context, the inventor has identified genes of which expressions are high in AGM-s3-A9 cell line which has the activity to support hematopoietic stem cells, and low or undetected in AGM-s3-A7 cell line which does not have the activity to support hematopoietic stem 25 cells, and has determined the activities to support hematopoietic stem cells, of cells in which these gene groups are highly expressed. As a result, the present

invention has been completed.

That is, the present invention provides the followings.

- (1) A DNA coding for a polypeptide of the
 5 following (A) or (B):
 - (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 23 and SEQ ID NO: 25; or
- (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- 15 (2) The DNA according to (1), which is a DNA of the following (a) or (b):
 - (a) a DNA which comprises a nucleotide sequence selected from the group consisting of the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 18, the nucleotide sequence of nucleotides 630 to 1358 of SEQ ID NO: 22, and the nucleotide sequence of nucleotides 132 to 506 of SEQ ID NO: 24; or
 - (b) a DNA which is hybridizable with a DNA comprising the nucleotide sequence as defined in (a) or a prove prepared from said DNA, under the stringent condition, and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic

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progenitor cells.

- (3) The DNA according to (2), the stringent condition is 6 x SSC, 5 x Denhardt, 0.5% SDS and 68°C (SSC: 3 M NaCl, 0.3 M sodium citrate; 50 x Denhardt: 1% BSA, 1% polyvinyl pyrrolidone, 1% Ficoll 400), or 6 x SSC, 5 x Denhardt, 0.5% SDS, 50% formamide and 42°C.
- (4) A expression vector which comprises the DNA of any one of (1) to (3) in such a manner that the DNA can be expressed.
- 10 (5) A cell into which the DNA of any one of (1) to (3) is introduced in such a manner that the DNA can be expressed.
 - (6) A polypeptide which is an expression product of the DNA of any one of (1) to (3), the polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- (7) The polypeptide according to (6), which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 23 and SEQ ID NO: 25, or an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence.
- (8) The polypeptide according to (6) or (7),
 25 which is modified with one or more modifying agents selected from the group consisting of polyethylene glycol (PEG), dextran, poly(N-vinyl-pyrrolidone),

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polypropylene glycol homopoymer, copolymer of polypropylene oxide/ethylene oxide, polyoxyethylated polyol and polyvinyl alcohol.

- (9) An monoclonal antibody which binds to the polypeptide of any one of (6) to (8).
 - (10) A method for supporting proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, comprising the step of co-culturing stromal cells in which a DNA coding for a polypeptide of the following (A) or (B) is expressed, with hematopoietic stem cells or progenitor cells,
 - (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or
 - (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
 - (11) The method according to (10), wherein the DNA is a DNA of the following (a) or (b):
- 25 (a) a DNA which comprises a nucleotide sequence selected from the group consisting of the nucleotide sequence of nucleotides 1 to 1671 of SEQ ID NO: 8, the



nucleotide sequence of nucleotides 1 to 1674 of SEQ ID

NO: 10, the nucleotide sequence of nucleotides 1 to 366

of SEQ ID NO: 12, the nucleotide sequence of nucleotides

84 to 1121 of SEQ ID NO: 14, the nucleotide sequence of

nucleotides 1 to 1035 of SEQ ID NO: 16, the nucleotide

sequence of nucleotides 1 to 444 of SEQ ID NO: 18, the

nucleotide sequence of nucleotides 1 to 444 of SEQ ID

NO: 20, the nucleotide sequence of nucleotides 630 to

1358 of SEQ ID NO: 22, the nucleotide sequence of

nucleotides 132 to 506 of SEQ ID NO: 24, the nucleotide

sequence of nucleotides 1 to 2487 of SEQ ID NO: 26, and

the nucleotide sequence of nucleotides 1 to 2496 of SEQ

ID NO: 28; or

- (b) a DNA which is hybridizable with a DNA comprising the nucleotide sequence as defined in (a) or a prove prepared from said DNA, under the stringent condition, and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- 20 (12) A method for supporting proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, comprising the step of culturing hematopoietic stem cells or progenitor cells in the presence of a polypeptide of the following (A) or (B), said polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells when the hematopoietic

stem cells or hematopoietic progenitor cells are cultured in the presence of the polypeptide,

(A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or

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- (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion
 10 of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- effect to support proliferation or survival of
 hematopoietic stem cells or hematopoietic progenitor
 cells, which comprises an effective amount of a
 polypeptide of the following (A) or (B), said
 polypeptide having an activity to support proliferation
 or survival of hematopoietic stem cells or hematopoietic
 progenitor cells when hematopoietic stem cells or
 hematopoietic progenitor cells are cultured in the
 presence of the polypeptide,
- (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23,



SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or

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(B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.

Terms used in this specification are defined as follows.

10 A hematopoietic stem cell is defined as a cell having totipotency, that is, ability to differentiate into all the cell lineages of the blood cells, and having a potency of self-renew with retaining the totipotency. A hematopoietic progenitor cell is defined as a cell which can differentiate a single cell lineage 15 of the blood cell or plural cell lineages but cannot differentiate into all of the cell lineages. A stromal cell is defined as a cell which can be co-cultured together with the hematopoietic stem cells to construct 20 a hematopoietic environment simulating in vivo hematopoietic environment in vitro. Cells derived from any origin can be used as long as the cells can be cocultured with the hematopoietic cells in vitro.

25 proliferate in *in vitro* culture environments and rapidly disappear. If the survival and proliferation of the erythrocyte progenitor cells are observed, continuous

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production of the erythrocyte progenitor cells is predicted to occur due to the survival and proliferation of the more immature hematopoietic stem cells or the hematopoietic progenitor cells. Therefore, in an assessment system of human hematopoietic stem cells, proliferation of hematopoietic stem cells or immature hematopoietic progenitor cells can be determined by using the survival and proliferation of the erythrocyte progenitor cells (BFU-E, CFU-E, and CFU-E mix) as an index.

Brief Explanation of the Drawings

- Fig. 1 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-s3 subclone A9, A7, or D11 cells for two weeks.
- Fig. 2 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-s3 subclone A9, A7, or OP9 cells for two weeks.
- Fig. 3 shows time course of donor derived lymphoid lineage cells or myeloid lineage cells reconstitution in irradiated recipient mice that received the hematopoietic stem cells co-cultured with stromal cells.
 - Fig. 4 shows proliferation statuses of hematopoietic



stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-2 is highly expressed (A9/SCR-2), AGM-S3-A9 cells into which a control vector is introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

Fig. 5 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A7 cells in which a gene SCR-2 is highly expressed (A7/SCR-2), AGM-S3-A7 cells into which a control vector is introduced (A7/pMXIG) or AGM-S3-A7 cells (A7) for two weeks.

Fig. 6 shows time course of donor derived lymphoid

lineage cells or myeloid lineage cells reconstitution in

peripheral blood of irradiated recipient mice that

received the hematopoietic stem cells co-cultured with

AGM-S3-A7 cells in which a gene SCR-3 is highly

expressed (A7/SCR-3), AGM-S3-A7 cells into which a

control vector is introduced (A7/pMXIG) or AGM-S3-A7

cells.

Fig. 7 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-4 is highly expressed (A9/SCR-4), AGM-S3-A9 cells into which a control vector is introduced

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(A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

Fig. 8 shows time course of donor derived lymphoid lineage cells or myeloid lineage cells reconstitution in peripheral blood of irradiated recipient mice that received the hematopoietic stem cells co-cultured with AGM-S3-A7 cells in which a gene SCR-5 is highly expressed (A7/SCR-5), AGM-S3-A7 cells into which a control vector is introduced (A7/pMXIG) or AGM-S3-A7 cells.

Fig. 9 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-6 is highly expressed (A9/SCR-6), AGM-S3-A9 cells into which a control vector is introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

Fig. 10 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-7 is highly expressed (A9/SCR-7), AGM-S3-A9 cells into which a control vector is introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

25 Fig. 11 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture



of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-8 is highly expressed (A9/SCR-8), AGM-S3-A9 cells into which a control vector is introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

Detailed Description of the Invention

Hereafter, the present invention will be described in detail below.

The following genes are those identified as genes of which expressions are high in AGM-s3-A9 cell line which has the activity to support hematopoietic stem cells, and low or undetected in AGM-s3-A7 cell line which does not have the activity to support hematopoietic stem

15 cells, and determined to have the activities to support hematopoietic stem cells, of cells in which these gene groups are highly expressed.

Gene SCR-2

The gene is the same gene as a mouse gene, Mus

musculus glypican-1 (GPC-1) of a GenBank accession

number AF185613.

The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 8. Only the amino acid sequence is shown in SEQ ID NO: 9.

The human amino acid sequence of GPC-1 is recorded

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in GenBank under an accession number P35052, and the human nucleotide sequence of GPC-1 is recorded in GenBank database under an accession number AX020122. It is predicted that the similar activity is detected in the human gene.

The nucleotide sequence of the gene from human and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 10. Only the amino acid sequence is shown in SEQ ID NO: 11.

20 Glypican is a major hepran sulfate proteoglycan existing on a cell surface, and have a characteristic structure such as cysteine rich globular domain, short glycosaminoglycan binding domain, glycosylphosphatidylinositol membrane binding domain. Six family genes from glypican-1 to glypican-6 have been found (J Biol Chem 1999 Sep 17;274(38):26968-77, Glypican-6, a new member of the glypican family of cell surface heparan sulfate proteoglycans. Veugelers M, De Cat B, Ceulemans H, Bruystens AM, Coomans C, Durr J, Vermeesch J, Marynen P, David G).

With respect to biological activities of GPC-1,
there are a number of reports: To regulate growth
stimulating activity of heparin binding growth factors
(fibroblast growth factor 2 (FGF2), heparin-binding EGFlike growth factor (HB-EGF)) to promote proliferation of
cancer cells showing autocrine proliferation by
stimulation by the growth factors (J Clin Invest 1998



Nov 1; 102(9):1662-7.3, The cell-surface heparan sulfate proteoglycan glypican-1 regulates growth factor action in pancreatic carcinoma cells and is overexpressed in human pancreatic cancer., Kleeff J, Ishiwata T, Kumbasar A, Friess H, Buchler MW, Lander AD, Korc M).

To bind HGF (hepatocyte groth factor) to promote reactivity with cytokines, of antigen-specific B cells. To participate in association of a cell with an adhesive molecule to involve in invasion of the cell (J Biol Chem 10 1998 Aug 28;273(35):22825-32, Heparan sulfate proteoglycans as adhesive and anti-invasive molecules. Syndecans and glypican have distinct functions., Liu W, Litwack ED, Stanley MJ, Langford JK, Lander AD, Sanderson RD). These findings show that GPC-1 involves 15 in activity expression of various cell-stimulating factors. Also, there is a report that expression of the glypican family gene in bone marrow is confirmed (Biochem J 1999 Nov 1;343 Pt 3:663-8, Expression of proteoglycan core proteins in human bone marrow stroma., 20 Schofield KP, Gallagher JT, David G reports, it is not described about effects of GPC-1 on hematopoietic stem cells or hematopoietic progenitor cells.

25 Gene SCR-3

The gene is the same gene as mouse genes, Mus

musculus chemokine MMRP2 mRNA of a GenBank accession

number U15209, Mus musculus C10-like chemokine mRNA of U19482 and mouse macrophage inflammatory protein-lgamma mRNA of U49513.

The nuclotide sequence of the gene from mouse and
the amino acid sequence deduced from the nucleotide
sequence are shown in SEQ ID NO: 12. Only the amino
acid sequence is shown in SEQ ID NO: 13.

Gene SCR-4

The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 14. Only the amino acid sequence is shown in SEQ ID NO: 15.

It has been found that the sequence has a high homology to *Homo sapiens* clone 25077 mRNA of a GenBank accession number AF131820, and that it is considered to be a mouse ortholog. This sequence is described in WO 00/66784.

The nuclotide sequence of the gene from human and
the amino acid sequence deduced from the nucleotide
sequence are shown in SEQ ID NO: 16. Only the amino
acid sequence is shown in SEQ ID NO: 17.

Gene SCR-5

The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 18. Only the amino

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acid sequence is shown in SEQ ID NO: 19.

It has been found that the sequence has a high homology with Homo sapiens esophageal cancer related gene 4 portein (ECRG4) mRNA of a GenBank accession number AF325503, and that it is considered to be a mouse ortholog of AF325503.

The nuclotide sequence of the gene from human and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 20. Only the amino acid sequence is shown in SEQ ID NO: 21.

Gene SCR-6

The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 22. Only the amino acid sequence is shown in SEQ ID NO: 23.

Gene SCR-7

The nuclotide sequence of the gene from mouse and
the amino acid sequence deduced from the nucleotide
sequence are shown in SEQ ID NO: 24. Only the amino
acid sequence is shown in SEQ ID NO: 25.

Gene SCR-8

The gene is the same gene as Mus musculus mRNA for ADAM23 of a GenBank accession number AB009673.

The nuclotide sequence of the gene from mouse and

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the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 26. Only the amino acid sequence is shown in SEQ ID NO: 27.

The sequence has a high homology with a sequence described by JP 11155574-A and the sequence described by JP 11155574-A is considered to be a human ortholog.

The nuclotide sequence of the gene from human and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 28. Only the amino acid sequence is shown in SEQ ID NO: 29.

Polypeptides which are products of these genes have an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells. The expression that a polypeptide has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells means that proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells is supported in the presence of the polypeptide or in the presence of stroma cells expressing the polypeptide.

Therefore, the present invention provides use of the polypeptides and DNAs encoding the polypeptides and novel polypeptides among the polypeptides and DNAs encoding the novel polypeptides.

A stem cell proliferation-supporting factor which is



a polypeptide encoded by the DNA can be produced by introducing the DNA into a suitable host to prepare a transformant cell, and allowing the DNA to be expressed in the transformant cell.

The DNA may encode the above described factors which have amino acid sequences including substitution, deletion or insertion of one or several amino acids, as long as the activity of the stem cell proliferation—supporting factor to be encoded is not lost. DNAs encoding substantially equivalent polypeptides to this stem cell proliferation—supporting factor can be obtained by modifying the nucleotide sequences so as to include substitution, deletion, insertion, addition, or inversion of amino acid residues in a specific region using site-directed mutagenesis.

The DNAs including the above described mutation can be expressed in appropriate cells and the activity to support hematopoietic stem cells, of the expressed products can be examined, so that the DNAs encoding the polypeptide having functions which are substantially equivalent to this stem cell proliferation-supporting factor are obtained. In addition, the DNAs encoding substantially equivalently active protein as this stem cell proliferation-supporting factor can be obtained by isolating DNAs which hybridize with DNAs including, for example, the nucleotide sequence as described in SEQ ID NO: 8, 10, 12, 14, 16, 18, 20, 22, 24, 26 or 28 from the

cells having the DNA, or probes prepared from these DNAs under the stringent condition; and which encode proteins possessing the activity to support hematopoietic stem cells. The length of the probe is usually 30 to 1000 5 nucleotides. The stringent condition is, for example, one in which DNAs having homology (determinable with homology search in the compare function of DNASIS version 3.7 (Hitachi Software Engineering)) at not less than 70%, preferably at not less than 80%, are 10 hybridized each other and DNAs having less homology than those are not hybridized each other. The above described stringent condition may be 6 \times SSC, 5 \times Denhardt, 0.5% SDS, 68°C (SSC; 3 M NaCl, 0.3 M sodium citrate) (50 × Denhardt; 1% BSA, 1% polyvinyl 15 pyrrolidone, 1% Ficoll 400) or 6 x SSC, 5 x Deanhardt, 0.5% SDS, 50% Formamide, 42°C, or the like.

Microorganisms such as Escherichia coli and yeast, culture cells derived from animals or plants, and the like are used for host cells for expressing the DNA.

- 20 Preferably, culture cells derived from mammals are used as the host cells. In the case that prokaryotic cells are used as the host cells, the expression is preferably performed in a condition in which a signal peptide region is replaced with a leader sequence suitable for the prokaryotic cells such as β-lactamase (bla),
- alkaline phosphatase (phoA), and outer membrane protein

 A (ompA) and the like, or in a form in which a



methionine residue is added to the N-terminal site of the mature protein.

The introduction of the DNA to the host cell can be carried out by, for example, incorporating the DNA into a vector suitable for the host in an expressible form, and introducing the resultant recombinant vector to the host cell.

Examples of the culture cells derived from mammals include CHO cell, 293 cell, COS7 cell, and the like.

- 10 Gene expression regulatory sequence such as a promoter to express the DNA may be originated from the gene itself, or may be derived from other genes such as cytomegalovirus promoter and elongation factor 1 promoter and the like.
- include plasmid vectors, retrovirus vectors, adenovirus vectors (Neering, S.J., Blood, 88: 1147, 1996), herpes virus vectors (Dilloo, D., Blood, 89: 119, 1997), HIV vectors, and the like.
- In order to introduce the recombinant vector into culture cells, the conventional methods which are usually employed for transformation of culture cells such as calcium phosphate transfection, the liposome method, the DEAE dextran method, the electroporation method and the microinjection method are employed.

The polypeptides of the present invention also comprise polypeptides having amino acid sequences in

which one or several amino acids are substituted, deleted or inserted in the amino acid sequence represented in SEQ ID NO: 9, 11, 13, 15, 17, 19, 21, 23, 25, 27 or 29, and having activity to support

- hematopoietic stem cells in addition to the polypeptides having the amino acid sequence represented in SEQ ID NO: 9, 11, 13, 15, 17, 19, 21, 23, 25, 27 or 29. That is, even if mouse and human stem cell proliferation—supporting factors are modified by substitution,
- deletion, insertion or the like, polypeptides holding essential functions as a stem cell proliferation—supporting factor can be considered to be substantially equivalent to the stem cell proliferation—supporting factor.
- 15 These modified stem cell proliferation-supporting factors can be obtained by treating DNA encoding the stem cell proliferation-supporting factor or host cells including the above mentioned DNA with a mutagen, or by mutating the above mentioned DNA so as to substitute,

 20 delete, or insert an amino acid at a specific site using
- delete, or insert an amino acid at a specific site using site-directed mutagenesis. The residual of the activity to support the hematopoietic stem cells in the obtained mutant polypeptide is confirmed by transferring hematopoietic stem cells cultured in the presence of the
- 25 mutant polypeptides into irradiated mice, and monitering peripheral hematological cellularity over time, as in the examples described below.



As for the amino acid deletion, the polypeptide may be a fragment which lacks an amino acid sequence at the N-terminal end and/or the C-terminal end. The fragment lacking the amino acid sequence at the N-terminal end and/or the C-terminal end can be obtained by a usual method, and the hematopoietic stem cell-supporting activity of the fragment can be determined by a similar way to that described with respect to the mutated polypeptide. In particular, if there is a portion 10 predicted as a signal sequence or a transmembrane region in the amino acid sequence, a fragment having the hematopoietic stem cell-supporting activity is predicted by using it as an index. For example, a protein encoded by human SCR-8 is a transmembrane protein of type I passing through the membrane once, and it is therefore 15 predicted that even if it made to be a soluble protein lacking the transmembrane region, it has the activity to support to proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells. The transmembrane region can be predicted with a known 20 program based on the amino acid sequence. For example, if it is predicted with a program called PSORT II (available through the Internet, URL: http://psort.nibb.ac.jp/index.html), the transmembrane region is amino acids at positions 790 to 806 in SEQ ID 25 NO: 29, and it is predicted that even if a fragment up to position 789, the fragment has activity to support

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proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.

Since the nucleotide sequences of the above described DNAs have been clarified by the present invention, the DNAs can be also obtained by isolating the corresponding DNAs from mouse or human cDNA or chromosome DNA libraries using PCR in which the oligonucleotides prepared based on these nucleotide sequences are used as primers or using hybridization in which the oligonucleotides prepared based on these nucleotide sequences are used as probes.

In order to complete the present invention, the DNAs of the present invention have been isolated from cDNA library of AGM-s3-A9 cells which are a mouse stromal cell line having the activity to support the hematopoietic stem cells, using SBH (Sequencing By Hybridization) method (Drmanac, S., Nat. Biotechnol., 16. 54, 1998; Drmanac, R., Methods. Enzymol., 303, 165, 1999) as described below. The mouse stromal cell lines having the activity to support the hematopoietic stem cells can be obtained using the method disclosed in W099/03980 or from Cell Bank of Institute of Physical and Chemical Research (RIKEN) or ATCC.

An outline of SBH method will be described below.

25 Probes having eight or nine nucleotides whose sequences are different from each other are prepared. When the nucleotide sequences corresponding to those of the probe



exist in a targeted gene, the probes can hybridize with the gene. The hybridization can be easily detected with utilization of radio isotope- or fluorescence-labelled probes. Each clone in the library is picked up, and blotted on a membrane. Then, the repeated . 5 hybridizations are performed with the each of above described probes, so that one can identify the combination of probes that hybridize to each clone. Since the combination of hybridized probes depends on 10 genes, the combination of probes which hybridize to an identical gene is the same. That is, the same gene can be identified as one group (cluster) according to the the combination of the hybridized probes. The number of clones of each gene in the cDNA library can be determined by classifying each clone in the library 15 based on patterns of the hybridized probes and counting the classified clones. Thus, frequency of expression of each gene in the library can be determined.

activity to support the hematopoietic stem cells and from cells not having the activity. Clustering is performed for the cDNA libraries. Statuses of expressed genes among cells are compared, so that the genes highly expressed with specificity to the supporting cells are selected. The expression statuses of these genes in each of above described cells are further examined by Northern blot analysis, so that genes which are highly



expressed in the cells having the activity to support the hematopoietic stem cells are obtained.

The above mentioned genes are the genes which are highly expressed with specificity to the supporting cells and which are obtained through the above described process. Full-length genes have been cloned from the cDNA library derived from AGM-s3-A9 cell.

Further, in order to determine an ability of gene products to support hematopoiesis, a gene fragment including gene ORF was transferred into stromal cells 10 using a retrovirus vector, and the change in the activity to support the hematopoietic stem cells of the stromal cells was determined. Specifically, after the stromal cells into which the gene was not introduced or into which a control vector was introduced and those 15 into which the gene was introduced were each co-cultured with the mouse hematopoietic stem cells, the hematopoietic cells were transplanted into irradiated mice. The engraftment of the co-cultured hematopoietic cells in recipient mice were examined. As a result, the 20 mice into which the hematopoietic stem cells co-cultured with the gene-introduced cells were transplanted, showed increased chimerism after the transplantation compared with co-culture with the cells into which the gene was not introduced. This result shows that in the gene-25 expressed stromal cells, an activity to support the proliferation or survival of the hematopoietic stem



cells or the hematopoietic progenitor cells is increased or imparted. As a result, it has become evident that expression of the above described genes has a function to increase the above described activity in the stromal cells or impart the activity to the stromal cells.

Therefore, it is revealed that products of the genes affect hematopoietic stem cells or hematopoietic progenitor cells to show an activity to support the survival or the proliferation thereof, or affect stromal cells to show an activity to increase an activity to support the hematopoietic stem cells therein or impart the activity thereto.

The polypeptides of the present invention can be used as a medicine to proliferate or support human hematopoietic stem cells or human hematopoietic progenitor cells when they affect hematopoietic stem cells or hematopoietic progenitor cells to show an activity to support survival or proliferation thereof, in other words, the polypeptides have an activity to 20 support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor cells if the hematopoietic stem cells or the hematopoietic progenitor cells are cultured in the presence of the polypeptides. The pharmaceutical composition can be used for supporting proliferation or survival of human 25 hematopoietic stem cells or human hematopoietic progenitor cells in vitro. For hematopoietic stem cell



transplantation therapies such as peripheral blood stem cell transplantation and cord blood stem cell . transplantation, a sufficient amount of the hematopoietic stem cells sometimes cannot be collected and the transplantation may not be performed. Even if 5 the enough amount of the stem cells can not be collected, the enough amount of the hematopoietic stem cells can be obtained and transplanted by amplification of the hematopoietic stem cells in vitro using this polypeptides. That is, the hematopoietic stem cells can 10 be amplified without differentiation by culturing the hematopoietic stem cells in culture medium including these polypeptides. It may be considered the hematopoietic stem cells are able to be amplified more efficiently with addition of a variety of cytokines to 15 the medium.

When the hematopoietic stem cells or the
hematopoietic progenitor cells are cultured in the
medium including the polypeptides of the present
invention, the hematopoietic stem cells or the
hematopoietic progenitor cells used may be isolated one
of these cell types alone or may be both of the cell
types. In addition, the cells may include at least the
hematopoietic stem cells or the hematopoietic progenitor
cells, and include other hematopoietic cells. Further,
it can be used a fraction containing hematopoietic stem
cells or progenitor cells fractionated from the cell

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population that contain the hematopoietic stem cells or progenitor cells.

Examples of sources of the hematopoietic stem cells and the hematopoietic progenitor cells in the method of the present invention include a fetal liver, bone marrow, fetal bone marrow, peripheral blood, the peripheral blood from persons whose stem cells are mobilized by administration of cytokines and/or antitumor drugs, cord blood, and the like of mammals such as human and mouse and the like. Any sources may be used as long as the tissue includes the hematopoietic stem cells.

A culture method using petri dishes and flasks for culture may be employed to culture the hematopoietic stem cells or the hematopoietic progenitor cells. The cultivation of the hematopoietic stem cells and/or progenitor cells may be improved by mechanically controlling medium composition, pH, and the like, and using a bioreactor capable of high density cultivation (Schwartz, Proc. Natl. Acad. Sci. U.S.A., 88: 6760, 1991; Koller, M.R., Bio/Technology, 11: 358, 1993; Koller, M.R., Blood, 82: 378, 1993; Palsson, B.O., Bio/Technology, 11: 368, 1993).

The stromal cells in which DNAs encoding the polypeptide of the present invention can be obtained as described with respect to the expression of the DNAs.

The co-culture of the stromal cells and the hematopoietic cells can be performed simply after the

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collection of the bone marrow cells without further separation. Furthermore, co-culture can be performed by separating components such as stromal cells, hematopoietic cells and other cell populations from collected bone marrow and combining them with the hematopoietic cells and stromal cells which are not from the individual from which the bone marrow is cllected. Furthermore, after stromal cells are cultured to grow to the stromal cells, hematopoietic cells can be added to perform co-culture with the hematopoietic stem cells. 10 At this time, cell stimulating factors can added to the culture system of stromal cells to more effectively support proliferation and survival. Concrete examples of the cell stimulating factor include a growth factor which is typically a cytokine such as SCF (stem cell 15 factor), IL-3 (interleukin 3), GM-CSF (granulocyte/macrophage colony-stimulating factor), IL-6 (interleukin 6), TPO (thrombopoietin), G-CSF (granulocyte colony-stimulating factor), TGF-b (transforming growth factor-b), MIP-la (Davatelis, G., J. 20 Exp. Med. 167: 1939, 1988); a differentiation and proliferation control factor such as hematopoietic hormones such as EPO (erythropoietin), chemokine, Wnt gene product, and notch ligand; and a development control factor. 25

In addition, when the polypeptide of the present invention affects hematopoietic stem cells or

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hematopoietic progenitor cells to show an activity to support survival or proliferation thereof, in other words, the polypeptide has an activity to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor cells if the hematopoietic stem cells or the hematopoietic progenitor cells are cultured in the presence of the polypeptide, the proliferation and the survival of the hematopoietic stem cells or the hematopoietic progenitor cells can be retained by allowing the recombinant polypeptide of the present invention alone or in combination with the cell stimulating factors to affect hematopoietic stem cells or hematopoietic progenitor cells, without stromal cells. Examples of the cell stimulating factors used in this case are above described cell stimulating factors and the like.

Medium used for the culture is not specially restricted as long as the proliferation or the survival of the hematopoietic stem cells or the hematopoietic progenitor cells is not harmed. Preferable media are, for example, MEM-α medium (GIBCO BRL), SF-02 medium (Sanko Junyaku), Opti-MEM medium (GIBCO BRL), IMDM medium (GIBCO BRL), and PRMI1640 medium (GIBCO BRL). A culture temperature is usually ranging from 25 to 39°C, and preferably ranging from 33 to 39°C. Examples of additives to the medium are fetal bovine serum, human serum, horse serum, insulin, transferrin, lactoferrin,



ethanolamine, sodium selenite, monothiolglycerol, 2-mercaptoethanol, bovine serum albumin, sodium pyruvate, polyethylene glycol, a variety of vitamins, and a variety of amino acids. A concentration of CO₂ is usually ranging from four to six percent, and preferably five percent.

Since the hematopoietic stem cells can differentiate into all the hematopoietic cell lineages, the hematopoietic stem cells can be amplified and differentiated into a specific cell type in vitro, and then the specific cells can be transplanted. For example, when erythrocytes are necessary, after the cultivation of the patient's stem cells to amplify them, the hematopoietic cells whose main component is the erythrocyte can be artificially produced using an erythrocyte differentiation induction-promoting factor such as EPO.

The hematopoietic stem cells or the hematopoietic progenitor cells cultured using the polypeptides of the present invention can be used as a graft for blood cell transplantation replacing the conventional bone marrow transplantation or cord blood transplantation.

Transplantation of the hematopoietic stem cells is superior to the conventional blood cell transplantation therapy, since the engraftment can last semipermanently.

The transplantation of the hematopoietic stem cells can be employed as therapy for a variety of diseases in

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addition to combination therapy with total body X-ray irradiation therapy or advanced chemotherapy for leukemia. For example, when therapy accompanied with myelosuppression as an adverse reaction, such as chemotherapy, radiation therapy, and the like is ∷ 5 performed for the patient with solid cancer, the patient can get benefit of early recovery and stronger chemotherapy than the conventional one can be performed to improve the therapeutic effect of the chemotherapy by collecting the bone marrow before the therapy, allowing 10 the hematopoietic stem cells or the hematopoietic progenitor cells to be amplified in vitro and returning the amplified cells to the patient after the therapy. In addition, by allowing the hematopoietic stem cells or the hematopoietic progenitor cells obtained according to the present invention to be differentiated into a variety of hematopoietic cells and transplanting these cells into a patient with hypoplasia of a given hematopoietic cells, the patient's deficient status can 20 be improved. In addition, this therapy can improve dyshemopoietic anemia to develop anemia such as aplastic anemia caused by bone marrow hypoplasia. Furthermore, examples of diseases in which the transplantation of the hematopoietic stem cells according to the present 25 invention is effective include immunodeficiency syndrome such as chronic granulomatous disease, duplicated immunodeficiency syndrome, agammaglobulinemia, Wiskott-

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Aldrich syndrome, acquired immunodeficiency syndrome (AIDS), and the like, thalassemia, hemolytic anemia due to an enzyme defect, congenital anemia such as sicklemia, Gaucher's disease, lysosomal storage disease such as mucopolysaccharidosis, adrenoleukodegeneracy, a variety of cancers and tumors, and the like.

Transplantation of the hematopoietic stem cells may be performed in the same manner as the conventional bone marrow transplantation or cord blood transplantation other than the differences of the cells used.

The source of the hematopoietic stem cells which may be used for the above described hematopoietic stem cell transplantation is not restricted to the bone marrow, and the above described fetal liver, the fetal bone marrow, the peripheral blood, the peripheral blood with stem cells mobilized by administration of cytokines and/or antitumor drugs, the cord blood, and the like may be used.

The graft may be a composition including buffer

20 solution and the like in addition to the hematopoietic stem cells and the hematopoietic progenitor cells produced by the method according to the present invention.

The hematopoietic stem cells or the hematopoietic

25 progenitor cells produced according to the present
invention may be used for ex vivo gene therapy. Because
of the low frequency of recombination of target genes to



the chromosome because the stem cells are in the resting state, differentiation of the stem cells during the culture period, and the like, the gene therapy to the hematopoietic stem cells has been hard to be established. :5 However, the present invention can amplify the stem cells without differentiation, so that efficacy of gene transfer is expected to be remarkably improved. In this gene therapy, a foreign gene (a gene for therapy) is transferred into the hematopoietic stem cells or the hematopoietic progenitor cells, and then the obtained 10 gene-transferred cells are used. The foreign gene to be transferred is appropriately selected according to disease. Examples of diseases in which the target cells of the gene therapy are the hematopoietic cells include 15 immunodeficiency syndrome such as chronic granulomatous disease, duplicated immunodeficiency syndrome, agammaglobulinemia, Wiskott-Aldrich syndrome, acquired immunodeficiency syndrome (AIDS), and the like, thalassemia, hemolytic anemia due to an enzyme defect, congenital anemia such as sicklemia, Gaucher's disease, 20 lysosomal storage disease such as mucopolysaccharidosis, adrenoleukodegeneracy, a variety of cancers and tumors, and the like.

A usual method used for transfer of a gene into

25 animal cells is employed for the transfer of the gene
for the therapy into the hematopoietic stem cells or the
hematopoietic progenitor cells. Examples include a

method using a vector for animal cells derived from virus utilized for a gene therapy such as retrovirus vectors such as Moloney mouse leukemia virus, adenovirus vectors, adeno-associated virus (AAV) vectors, herpes 5 simplex virus vectors, and HIV vectors (with respect to a vector for gene therapy, see Verma, I.M., Nature, 389: 239, 1997); calcium phosphate transfection, DEAE-dextran transfection, electroporation, the liposome method, the lipofection method, the microinjection method, and the like. Among them, the method using the retrovirus 10 vector, the adeno-associated virus vector, or the HIV vector is preferable, since permanent expression of a gene is expected due to insertion into the chromosome DNA of a target cell.

15 For example, the adeno-associated virus (AAV) vector can be prepared as follows. First, a vector plasmid in which a gene for therapy is inserted into ITR (inverted terminal repeat) at both ends of wild-type adenoassociated virus DNA and a helper plasmid for 20 supplementing virus proteins are transfected into 293 cell line. Next, adenovirus as helper virus is infected, so that virus particles including the AAV vector are produced. Alternatively, instead of adenovirus, a plasmid which expresses adenovirus gene having helper 25 function may be transfected. The hematopoietic stem cells or the hematopoietic progenitor cells are infected with the obtained virus particles. Preferably,



appropriate promoter, enhancer, insulator and the like are inserted into the upstream region of the target gene in the vector DNA, so that the expression of the gene is regulated. When a marker gene such as a drug resistant gene is used in addition to the gene for therapy, cells into which the gene for therapy are transferred are easily selected. The gene for therapy may be a sense gene or an antisense gene.

A composition for gene therapy may include a buffer solution and a novel active ingredient and the like in addition to the hematopoietic stem cells or the hematopoietic progenitor cells by the method according to the present invention.

A vector for gene therapy can be produced by incorporating the DNA of the present invention in an 15 expression vector using a usual method. This vector for gene therapy is useful to treat diseases which need survival and proliferation of the human hematopoietic stem cells. That is, the vector for gene therapy is transferred into the hematopoietic stem cells and the 20 cells are cultured in vitro, so that the hematopoietic stem cells or the hematopoietic progenitor cells can proliferate dominatingly. The proliferation of hematopoietic stem cells in vivo can be caused by returning these hematopoietic stem cells thus treated. 25 The proliferation of hematopoietic stem cells in vivo can significantly promoted by introducing this vector

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for gene therapy into the body. Alternatively, the bone marrow cells derived from a patient are cultured as it is and this vector for gene therapy is transferred thereto, so that the hematopoietic stem cells or the hematopoietic progenitor cells can be proliferated in a culture system. Alternatively, this vector for gene therapy is transferred into the stromal cells and mesenchaymal stem cells obtained by isolating and culturing stromal cells from the bone marrow, so that the activity to support the hematopoietic stem cells can be added or increased.

As shown in Examples, since it is possible that by introducing the DNA of the present invention into the stromal cells without the activity to support the

15 hematopoietic stem cells, this activity can be imparted, stromal cells having the activity to support the hematopoietic stem cells can be prepared by gene transfer to stromal cells derived from human or mouse.

The stromal cells expressing the DNA of the present

20 invention and the hematopoietic stem cells or the hematopoietic progenitor cells are co-cultured, so that the hematopoietic stem cells or the hematopoietic stem cells or the hematopoietic progenitor cells can survive and proliferate so as to be useful for a variety treatment.

25 Since the hematopoietic stem cells or the hematopoietic progenitor cells can survive and proliferate by expression of the DNA of the present

invention in the stromal cell, an activity to support
the hematopoietic stem cells of the stromal cells can be
determined using the expression of the DNA of the
present invention as an index. The expression of the
DNA of the present invention in the stromal cells can be
confirmed using an antibody against a polypeptide
encoded by the DNA of the present invention. Also, PCR
primers can be prepared based on nucleotide sequences,
and RNA is prepared from the stromal cells of interest,
and RT-PCR is performed, so that the expression of the
DNA of the present invention can be confirmed. The
antibody will be described below.

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The pharmaceutical composition of the present invention can be administered to human. The 15 pharmaceutical composition having an activity to proliferate or to support the human hematopoietic stem cells or the hematopoietic progenitor cells can be produced by mixing medically acceptable diluent, stabilizer, carrier, and/or other additives with the 20 polypeptides of the present invention. At this time, in order to increase the stability of the protein in vivo, the polypeptides of the present invention may be modified by a modifying agent. Examples of the modifying agent include polyethylene glycol (PEG), 25 dextran, poly(N-vinyl-pyrrolidone), polypropylene glycol homopolymer, polypropylene oxide/ethylene oxide copolymer, polyoxyethylated polyol, polyvinyl alcohol,

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and the like. The modification of the protein with PEG can be performed by, for example, a method in which activated ester derivatives of PEG is reacted with the protein, a method in which aldehyde derivatives at the terminal portion of PEG is reacted with the protein in the presence of a reducing agent, and the like.

Japanese Patent Application Laid-Open No. 10-510980 discloses such protein modification in detail.

When the pharmaceutical composition of the present invention is administered to human, recovery from hematological suppression due to an adverse drug reaction of carcinostatics; early recovery of hematopoietic cells at bone marrow transplantation, peripheral blood stem cell transplantation, or cord blood transplantation; and recovery of hematopoietic function at pancytopenia such as aplastic anemia (AA) and myelodysplastic syndrome (MDS) are expected.

The antibodies of the present invention react specifically to the above described polypeptides of the present invention. The antibodies of the present invention may be monoclonal antibodies or polyclonal antibodies as long as they react specifically to the above described polypeptides.

The antibodies of the present invention can be

25 prepared according to usual methods. For example, the

antibodies can be prepared either *in vivo* method in

which animals are additionally immunized by an antigen



together with adjuvant once or several times at an interval of several weeks or in vitro method in which immune cells are isolated and sensitized in an appropriate culture system. Examples of immune cells which can produce the antibodies of the present invention include splenic cells, tonsillar cells, lymph gland cells, and the like.

The whole polypeptide according to the present invention is not necessarily used as an antigen. A part 10 of this polypeptide may be used as an antigen. When the antigen is a short peptide, particularly, a peptide made of about 20 amino acid residues, it may be used by binding it to a carrier protein having high antigenicity such as keyhole lympet hemocyanin or bovine serum albumin using chemical modification and the like. 15 Alternatively, the antigen may be used by covalently binding it to peptide having branching skeleton such as lysine core MAP peptide instead of the carrier protein (Posnett et al., J. Biol. Chem., 263, 1719-1725, 1988; Lu et al., Mol. Immunol., 28, 623-630, 1991; Briand et 20 al., J. Immunol. Methods, 156, 255-265, 1992).

Examples of adjuvant include Freund's complete
adjuvant, Freund's incomplete adjuvant, aluminum
hydroxide gel, and the like. Antigen-given animals are,
for example, mouse, rat, rabbit, sheep, goat, chicken,
bovine, horse, guinea pig, hamster, and the like. The
blood is collected from these animals and the serum is

separated. Then, immunoglobulin is purified from the serum using an ammonium sulfate precipitation method, anion exchange chromatography, protein A chromatography, or protein G chromatography to obtain polyclonal antibodies.

With respect to chicken, antibodies can be purified from an egg. Monoclonal antibodies can be prepared by purification from supernatant of culture of hybridoma cells which are made by fusion of the immune cells sensitized in vitro, or immune cells from the above 10 described animals with parent cells capable of cultivation, or ascites from animals which received intraperitoneal administration of hybridoma cells. Examples of parent cells include X63, NS-1, P3U1, X63.653, SP2/O, Y3, SK0-007, GM1500, UC729-6, HM2.0, 15 NP4-1 cell lines, and the like. Preparation may be performed by cultivating the immortalized antibodyforming cells obtained by sensitization in vitro, or infection of a proper virus such as EB virus to the 20 immune cells of the above described animals.

In addition to these cell engineering methods, the antibodies can be obtained using gene engineering methods. For example, the antibody gene obtained from the *in vitro* sensitized cells or immune cells derived from the above described animals is amplified by PCR (polymerase chain reaction) and isolated, and the amplified genes are transferred into microorganisms such

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as *E. coli* to produce the antibodies. Alternatively, the antibodies may be expressed on surfaces of phages as fused proteins.

By measuring polypeptides in vivo using the antibodies of the present invention, the relationship between the polypeptides and pathological status of a variety of diseases can be clarified. Moreover, the antibodies can be used for diagnosis and treatment of diseases, and efficient affinity purification of the polypeptides.

The present invention provides polypeptides having an activity to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor cells by effecting thereon, or an activity to impart an activity to support the hematopoietic stem cells to stromal cells by effecting thereon, and also provides DNAs encoding thereof. The polypeptides of the present invention can efficiently maintain the proliferation or the survival of the hematopoietic stem cells or the hematopoietic progenitor cells.

Best Mode for Carrying out the Invention

Hereafter, the present invention will be described in detail by reference to examples.

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Example 1 Preparation of fragment of gene which is specifically expressed in hematopoietic stem cell-

supporting cells

- (I) Preparation of stromal cell line derived from mouse AGM
- 10 existence of a vaginal plug was observed were transferred to other cages and kept. The day when the existence of the vaginal plug was observed was defined to be the 0.5th day of pregnancy. On the 10.5th day of the pregnancy, after mouse was sacrificed by cervical
- dislocation, fetuses were extirpated. Isolation of AGM regions was performed according to the method by Godin et al. (Godin, I., Proc. Natl. Acad. Sci. U.S.A., 92: 773-777, 1995) and the method by Medvinsky et al. (Medvinsky, A.L., Blood, 87: 557-565, 1996). The
- fetuses were placed in a culture dishes to which PBS(-)
 (phosphate buffered saline) (produced by Nissui Seiyaku)
 was added in a volume just sufficient to cover the
 fetuses. After the AGM regions were carefully excised
 so as not to include other regions under a stereoscopic
- 25 microscope, they were put in another 24-well culture dish (Nunc).
 - (2) Establishment of cell lines derived from AGM



One drop of MEM medium (Sigma) containing 10% FCS (Hyclone) was added to the AGM regions in the 24-well culture dish (Nunc), and AGM regions were cultured in an incubator overnight. The culture was performed in the MEM medium (Sigma) containing 10% FCS (Hyclone) at 37°C, in an atmosphere of 5% CO2, and at a humidity of 100%. When the cells of the AGM regions adhered to the culture dish due to overnight cultivation, two milliliters of MEM medium containing 10% FCS was further added. Stromal cells began to appear around the AGM region 10 tissue fragment after the continuous cultivation. After one-week cultivation, adhesive cells were separated by trypsin treatment (0.05% trypsin in PBS containing 0.53 mM EDTA (Gibco BRL) at 37°C for three to five minutes). The stromal cells were then washed twice with the medium, 15 and seeded on 6-well culture dish (Nunc). On the next day, the cells which did not adhere to the culture dish and the medium were removed, and then, fresh medium was added. Two weeks after transfer to the 6-well culture dish, cells were y-ray-irradiated at 900 Rad to 20 eliminate endogenous hematopoietic cells. An attempt of the direct cell cloning by limiting dilution from this culture system was made, but no cell proliferation was

observed and the cloning ended in failure. Then, after

cells were adapted so as to be able to proliferate from

25 the number of seeded cells in one well was increased and

a small number of cells, the cells were cloned by

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limiting dilution.

Specifically, the AGM was extirpated and cultured in the same manner as described above. The culture system two weeks after the γ-ray radiation was trypsinized (0.05% trypsin in PBS containing 0.53 mM EDTA at 37°C for three to five minutes) to suspend the cells, and the cells were seeded in a 24-well culture dish at 50 to 100 cells/well. After the culture was continued for three weeks, the cells were seeded in a 96-well culture dish (Nunc) by means of limiting dilution, at 0.3 cells/well. The cells which were grown from the well in which only one cell was seeded were allowed to enlarge culture. As a result, the cells were successfully cloned to obtain fibroblast-like cells and cobble stone-like cells.

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A CD34-positive cell fraction derived from the human cord blood was co-cultured with the fibroblast-like cells for two weeks to examine the presence of colony-forming cells during the culture. Colony-forming cells could not be found in the co-culture system with the fibroblast-like cells. Then, the similar examination was performed for seven cell clones showing the cobblestone-like form. Three clones having an activity to support proliferation of human hematopoietic stem cells were obtained and were named AGM-s1, AGM-s2, and AGM-s3.

25 (II) Preparation of hematopoietic stem cells from mouse bone marrow

Bone marrow was collected from a femur of C57BL/6-

Ly5.1 pep (eight- to ten-week age, and male) (the gift from Professor K. Nakauchi, University of Tsukuba), and suspended in PBS. After the mouse bone marrow mononuclear cells were concentrated by specific gravity centrifugation according to the usual method (S. Kouzu, Fundamental techniques for immunology, YODOSHA, 1995), the cells were suspended in staining buffer (PBS containing 5% FCS and 0.05% NaN₃), and a hematopoietic stem cell fraction was obtained as follows (Osawa, M. et al., Science 273: 242-245, 1996).

An FITC-conjugated anti-CD34 antibody, a phycoerythrin-conjugated anti-Sca-1 antibody, an allophycocyanin anti-c-Kit antibody (all purchased from Pharmingen) and six biotylated anti-differentiation antigen antibodies (CD45R, CD4, CD8, Gr-1, Terl19, and 15 CD11c, all purchased from Pharmingen) as molecularmarkers (Lin), were added to a suspension of the bone marrow mononuclear cells and incubated for 20 min on ice to cause reaction. After the cells were washed twice with staining buffer, CD34-negative, Sca-1-positive, c-20 Kit-positive, and Lin-negative cells were isolated on a cell sorter (FACS Vantage, Becton Dickinson). (III) Subcloning of mouse stromal cell line and determination of activity to support hematopoietic stem 25 cells of a variety of cell lines

- (1) Subcloning of mouse stromal cell line
- 1) Isolation of AGM-s3 subclone

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Stromal cell line AGM-s3 derived from AGM, which was subcultured in MEMa medium (GIBCO BRL) including inactivated 10% FCS (bovine fetal serum, Hyclone), was suspended in PBS containing 5% FCS (PBS-FCS). Clone sorting was performed in a 96-well culture dish (Falcon) at one cell/well using a cell sorter (FACS Vantage; Becton Dickinson). Among cells in the 96 wells, cultures of the cells which grew were expanded, so that thirteen kinds of AGM-s3 subclones were obtained. The activity to support the hematopoietic cells of these AGM-s3 subclones were examined.

2) Isolation of human cord blood CD34-positive stem cell The human cord blood was collected at normal delivery according to the criteria approved by Ethics committee of Kirin Beer Iyaku Tansaku Kenkyusho. The cord blood was collected using a heparin-added syringe so as not to coagulate. The heparin treated cord blood was overlaid on Lymphoprep (NYCOMED PHARMA), and mononuclear cells were separated by specific gravity centrifugation (at 400G, at room temperature, and for 30 minutes). Erythrocytes contaminated in the mononuclear cell fraction were lyzed by treatment with an ammonium chloride buffer solution (0.83% NH₄Cl-Tris HCl, 20 mM, pH 6.8) at room temperature for two minutes. After the mononuclear cells were washed with PBS-FCS, ten milligrams of human IgG was added thereto and the mixture was allowed to stand on ice for ten minutes.

Then, the cells were further washed with PBS-FCS. Biotinylated antibodies against the antigens specific to the human differentiated blood cells, that is, the antibodies against CD2, CD11c (purified from ATCC hybridoma), CD19 (Pharmingen), CD15, and CD41 (Leinco Technologies Inc.), and Glycophorin A (Cosmo Bio) were added thereto, and the mixture was allowed to stand on ice for 20 min. After washing with PBS-FCS, the cells were suspended in one milliliter of PBS containing 5% 10 FCS, 10 mM EDTA, and 0.05% NaN3 (PBS-FCS-EDTA-NaN3). Streptavidin-bound magnetic beads (BioMag. Per Septive Diagnostics) were added thereto, and the mixture was allowed to stand on ice for 40 min. The differentiated blood cells which expressed differentiation antigens . were removed using a magnetic separator (Dynal MPC-1 :15 Dynal). An FITC-labeled anti-CD34 antibody (Immunotech S.A., Marseilles, France) was added to the remaining differentiated blood cell antigen-negative cell fraction. After incubation on ice for 20 min., a CD34-positive fraction was recovered using a cell sorter. This cell 20 population was defined as a hematopoietic stem cell population derived from the human cord blood. 3) Co-culture of the human hematopoietic stem cells and AGM-s3 subclone

25 After 13 kinds of AGM-s3 subclones and stromal cell line MS-5 derived from the mouse bone marrow were each seeded in a 24-well culture dish (Falcon) at 1×10^4

cells/well, and cells were cultured in one milliliter of MEMα medium containing 10% FCS and allowed to grow until the cells covered all over the bottom surfaces of the wells. CD34-positive hematopoietic stem cells derived from the human cord blood were placed on the above 5 described stromal cells at 500 cells/well, and cocultured in one milliliter of MEMa medium containing 10% FCS. One week after the start of the co-culture, one milliliter of the same medium was further added. Two weeks after the start of the co-culture, the stromal 10 cells and the human blood cells were trypsinized (0.05% trypsin in PBS containing 0.5 mM EDTA (GIBCO BRL) at 37°C; standing for two to five min.) to simultaneously separate them from the culture dish. An activity to support the hematopoietic stem cells was determined with 15 a clonogenic assay.

4) Assessment of proliferation statuses of the hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay

The cells which proliferated in the above described co-culture system were appropriately diluted, and subjected to one milliliter of methylcellulose culture system to be analyzed. The analysis using the methylcellulose culture system was performed using a 6-well culture dish (Falcon) in MethoCult H4230 (Stem Cell Technologies Inc.) to which 10 ng/ml of human SCF, human IL-3, human IL-6, human G-CSF, and human TPO, and 2

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IU/ml of EPO were added. All of a variety of the above described hematopoietic factors were recombinants and pure. After two-week culture, developed colonies were observed under a microscope to count numbers of CFU-GM (granulocyte-macrophage colony-forming unit), BFU-E (erythroid burst forming unit), and CFU-E mix (erythrocyte mixed colony-forming unit).

Fig. 1 shows the result of two-week co-culture of the CD34-positive hematopoietic stem cells and the AGMs3 subclone A9, A7, or D11. As a result of the co-10 culture, A9 and D11 subclones among 13 kinds of AGM-s3 subclones supported proliferation of all three series of CFU-GM, BFU-E, and CFU-E mix. Especially, although BFU-E and CFU-E mix, that is, the progenitor cells of erythrocytes were hardly to be supported in usual, their 15 proliferations were observed in the co-culture system with A9 or D11 cells. The results showed that proliferation or maintenance of the hematopoietic stem cells or the hematopoietic progenitor cells occurred in 20 the co-culture with A9 or D11 cells and the progenitor cells of the erythrocyte were continuously supplied. In contrast, although cellular morphology of A7 was similar to that of A9, A7 did not support CFU-GM, BFU-E, and CFU-E mix.

25 5) Comparison of an activity to support the human hematopoietic stem cells between A9 and a stromal cell line OP9 derived from mouse fetus



Comparison of an activity to support the CD34positive hematopoietic stem cells derived from the human cord blood between AGM-s3 subclones A9 and A7, and a stromal cell line OP9 derived from mouse fetus (RCB1124, the Cell Development Bank of RIKEN) were performed with 5 CFU-GM, BFU-E, CFU-E and CFU-E mix as indexes, using the above described determination system. Fig. 2 shows the result of the two-week co-culture. In the A7 cell culture system, CFU-GM, BFU-E, and CFU-E were significantly decreased and CFU-E mix was completely 10 disappeared. In contrast, with OP9 cells, a variety of blood cell progenitor cells including CFU-E mix were supported, although the supporting ability was less than that of A9 cells. Therefore, it has been found that OP9 cells possess the activity to support the hematopoietic 15 stem cells.

(2) Assessment of activity to support the hematopoietic stem cells in a variety of cell lines

The above described stromal cell lines (AGM-s3-A9,

20 AGM-s3-A7, and AGM-s3-G1), 3T3Swiss (ATCC), OP9, and

NIH3T3 (ATCC) were seeded in a 24-well culture dish

(Falcon) at 5 × 10⁴ cells/well. The cell lines were

cultured in MEMa medium (GIBCO BRL) containing

inactivated 10% FCS (bovine fetal serum, Hyclone) for

25 one day and allowed to proliferate until the cells

covered all over the bottom surfaces of the wells. Then

the medium was replaced to one milliliter of fresh

medium, thirty cells of the mouse hematopoietic stem cells (derived from C57BL/6-Ly5.1) obtained in the above (II) were placed on this cell layer, and co-culture was started.

On seventh day of the cultivation, the cells were

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trypsinized (0.05% trypsin in PBS containing 0.5 mM EDTA (GIBCO BRL) at 37°C for two to five minutes) to separate and recover all the cells on the culture dish. The recovered whole cells of each cell line and 200,000 10 cells of whole bone marrow cells (derived from C57BL/6-Ly5.2 mouse, Charles River) were transplanted into C57BL/6-Ly5.2 mice (eight weeks age and male, Charles River) irradiated with X-ray at 8.5 Gy through the tail vein. After the transplantation, peripheral blood was collected from orbit at intervals, and the ratio in 15 number of cells derived from the C57BL/6-Ly5.1 prep mouse was determined with FACS. The peripheral blood was analyzed according to the usual method (S. Kouzu, Fundamental techniques for immunology, YODOSHA, 1995). 20 Three hundreds and fifty μL of distilled water was added to 50 µL of the peripheral blood, and the mixture was allowed to stand for 30 seconds so as to lyze the erythrocytes. Then, PBS at twice concentrations was added and the mixture was centrifuged to recover white blood cells. After the cells were washed once using the 25 staining buffer (PBS containing 5% FCS and 0.05% NaN3),

anti-CD16 antibody, anti-Ly5.1 (CD45.1) antibody labeled

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with FITC, anti-Gr-1 and anti-CD11c antibodies labeled with phycoerythrin, and anti-CD45R (B220) and anti-CD90 (Thy1) antibodies labeled with allophycocyanin (all of these were purchased from Pharmingen) were added. After these cells were allowed to stand for reaction in the ice bath for 30 minites, they were washed with the staining buffer and FACS analysis was performed.

Change in the number of cells capable of reconstitution during the hematopoietic stem cell culture was determined by calculating the proportions of Ly5.1-positive cells in the Gr-1- or CD11c-positive cells (myeloid cells) and Ly5.1-positive cells in the CD90- or CD45R-positive cells (lymphoid cells) in the peripheral blood at intervals after transplantation.

Fig. 3 shows the results. When the cells were cocultured with AGM-s3-A9 cells, OP9 cells, or 3T3Swiss cells, high chimerism of donor cells were maintained after the transplantation. Therefore, these stromal cells were considered to have a high activity to support the hematopoietic stem cells. In contrast, when the cells were co-cultured with AGM-s3-A7 cells, AGM-s3-G1 cells, or NIH3T3 cells, high chimerism derived from the transplanted cells was not observed. Therefore, these stromal cells were low in an activity to support the hematopoietic stem cells or the hematopoietic progenitor cells.

(IV) Identification of sequences of genes which



specifically express in hematopoietic stem cellsupporting cells

AGM-s3-A9 cells, AGM-s3-A7 cells and OP9 cells were each dissolved in 20 mL of ISOGEN (Nippon gene, Japan) and total RNAs were prepared according to the attachment. Messenger RNAs were prepared from one milligram of the total RNAs according to the protocol of the mRNA purification kit (Amersham Pharmacia, U.S.A.). cDNAs were synthesized from the mRNAs and cDNA libraries (hereinafter, also called as AGM-s3-A9 cDNA, AGM-s3-A7 10 cDNA and OP9 cDNA, respectively) were constructed using pSPORT1 (GIBCO Lifetech, U.S.A.). A clone harboring a cDNA fragment which highly expresses specifically to AGM-s3-A9 cells or OP9 cells compared with AGM-s3-A7 15 cells was obtained from the libraries with SBH method (Hyseq, U.S.A.). A nucleotide sequence of the obtained clone was determined using ABI377 DNA sequencer (Perkin Elmer, U.S.A.).

As a result, it has been found that expression of

genes comprising nucleotide sequences shown in SEQ ID

NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5,

SEQ ID NO:6, and SEQ ID NO:7, or parts thereof in AGM
s3-A9 or OP9 cells is higher than that in AGM-s3-A7

cells. These genes were named as SCR-2, SCR-3, SCR-4,

SCR-5, SCR-6, SCR-7 and SCR-8, respectively.

Example 2 Cloning of SCR-2 and activity determination

By searching GenBank database for the nucleotide sequence shown in SEQ ID NO: 1 with BLAST, it has been found that SCR-2 is the same gene as a mouse gene, Mus musculus glypican-1 (Gpc-1) of an accession number AF185613. The nuclotide sequence of ORF (Open Reading Frame) of SCR-2 and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 8. Only the amino acid sequence is shown in SEQ ID NO: 9.

The human nucleotide sequence of Gpc-1 is recorded in GenBank database under an accession number AX020122.

The nucleotide sequence of ORF of AX020122 and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 10. Only the amino acid sequence is shown in SEQ ID NO: 11.

Determination of the activity to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

(1) Construction of retrovirus vector for expression of 20 mouse SCR-2

Based on the nucleotide sequence of SCR-2 ORF, SCR-2Fsall and SCR-2Reco primers having the following nucleotide sequences were prepared, and PCR was performed using OP9 cDNA as a template.

25 SCR-2Fsal

CCGGTCGACCACCatggaactccggacccgaggctgg (SEQ ID NO: 30)



SCR-2Reco

CCGAATTCttaccgccacctgggcctggctgc (SEQ ID NO: 31)

An amplified fragment was digested with restriction enzymes EcoRI and SalI. After electrophoresis, a DNA fragment was purified using JETSORB (Genomed, Germany). The purified DNA fragment was ligated with pMX-IRES-GFP vector digested with EcoRI and XhoI (gift form Professor T. Kitamura, TOKYO UNIV. INST. OF MEDICAL SCIENCE, Japan). The pMX-IRES-GFP vector is a plasmid obtained by inserting sequences encoding IRES (Internal Ribosome 10 Entry Site) and GFP (Green Fluorescence Protein) into the retrovirus vector pMX. IRES (Internal Ribosome Entry Site) enables ribosome to access to the middle of the mRNA. Therefore, two genes can be expressed from 15 one mRNA by ligation of upward and downward genes separated by IRES in one transcription unit during the construction of an expression vector. With respect to the above-described plasmid, SCR-2 cDNA was inserted in the upward site and GFP (Green Fluorescence Protein) was 20 inserted in the downward site. Thus, the expression of SCR-2 could be monitored by detecting the expression status of GFP using FACS.

The obtained recombinant vector was introduced into E. coli DH5α, and was seeded on LB agar medium

containing 100 μg/ml of ampicillin, so that independent colonies were formed. After the isolated colony was cultured in 100 mL of LB medium containing 100 μg/ml of

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ampicillin, plasmid was purified using QIAGENtip100 (QIAGEN, U.S.A.). The sequence of the inserted gene was determined using a conventional method, so that the sequence was confirmed to be identical to the nucleotide sequence of SCR-2 ORF.

(2) Preparation of stromal cells highly expressing SCR-2
BOSC23 cells were seeded on a collagen type I-coated
60-mm dish (Asahi technoglass) at 2 × 10⁶ cells/dish,
and cultured in DMEM medium containing 10% FCS at 37°C,
under an atmosphere of 5% CO₂, and at a humidity of 100%.
Twelve to 18 hours after the start of the culture, the
medium was replaced by two milliliters of OPTI MEM
medium (GIBCO BRL).

About 3 μg of plasmid obtained by inserting SCR-2

into the above described pMX-IRES-GFP was added to 18 μl

of LIPOFECTAMINE Reagent (GIBCO BRL) diluted with 100 μl

of OPTI MEM medium, and the mixture was allowed to stand

at room temperature for 30 min. The prepared DNA

solution was added to the prepared BOSC23 cell culture

20 solution. After about five hours, two milliliters of

DMEM medium containing 20% FCS (GIBCO BRL) was added.

After about 24 hours, the medium was replaced by 4 ml of DMEM containing 10% FCS. Further, after about 48 hours, the culture medium was harvested. After the culture medium was filtrated through 0.45-µm filter, the filtrate was centrifuged at 1,200g for 16 hours and the supernatant was removed to obtain the virus

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precipitation.

AGM-s3-A7 or AGM-s3-A9 cells were cultured in one milliliter of MEMa medium containing 10% FCS (GIBCO BRL) on a 24-well culture dish (FALCON) at 1×10^4 cells/well. After 12 to 18 hours, the virus precipitation was suspended in one milliliter of MEMa medium containing 10% FCS, and the stromal cell culture medium was replaced by the virus suspension. Next, POLYBRENE (Sigma, SEQUA-BRENE) was added to be 10 µg/ml. After 10 the culture dish was centrifuged at 700g for 45 minutes, the cells were cultured at 37°C, under an atmosphere of 5% CO2, and at a humidity of 100%. After 48 hours, the medium was replaced by one milliliter of MEMa medium containing 10% FCS. After 24 hours, the cells were subcultured on a 6-well culture dish (FALCON) and cultured in three milliliters of MEMa medium containing 10% FCS. Forty-eight hours after the subculturing, GFP expression in the stromal cells was detected using a cell sorter (FACSVantage, Becton Dickinson) to indirectly confirm that not less than 80% of cells 20 expressed SCR-2.

Also, the same procedures were repeated by using pMX-IRES-GFP vector instead of the plasmid obtained by inserting SCR-2 into pMX-IRES-GFP to prepare stromal cells into which a control vector was introduced.

(3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing SCR-2, and determination

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of proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay

In the same manner as described in (III) (1) 3) to 4) of Example 1, AGM-s3-A9 or AGM-s3-A7 cells in which SCR-2 was highly expressed through retrovirus, AGM-s3-A9 or AGM-s3-A7 cells into which a control vector was introduced, or AGM-s3-A9 or AGM-s3-A7 cells were co-cultured with CD34-positive hematopoietic stem cells derived from human cord blood, and proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells are determined.

Fig. 4 shows results when the CD34-positive hematopoietic stem cells were co-cultured with AGM-S3-A9 cells in which SCR-2 was highly expressed (A9/SCR-2), 15 AGM-S3-A9 cells into which a control vector was introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks. Also, Fig. 5 shows results when the CD34positive hematopoietic stem cells were co-cultured with AGM-S3-A7 cells in which SCR-2 was highly expressed, 20 AGM-S3-A7 cells into which a control vector was introduced or AGM-S3-A7 cells for two weeks. As a result, by the co-culture with AGM-S3-A9 cells in which SCR-2 was highly expressed or AGM-S3-A7 cells in which SCR-2 was highly expressed, increases of BFU-E and CFU-C 25 were observed. Therefore, it has been revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9 or AGM-S3-



A7 increases by allowing SCR-2 to be highly expressed. From the results, it has been revealed that a gene product of SCR-2 has an activity to support survival or proliferation of hematopoietic stem cells or

hematopoietic progenitor cells or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

10 Example 3 Cloning of SCR-3 and activity determination

By searching GenBank database for the nucleotide sequence shown in SEQ ID NO: 2 with BLAST, it has been found that SCR-3 is the same gene as mouse genes, Mus musculus chemokine MMRP2 mRNA of an accession number U15209, Mus musculus C10-like chemokine mRNA of U19482 and mouse macrophage inflammatory protein-lgamma mRNA of U49513. The nuclotide sequence of SCR-3 ORF and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 12. Only the amino acid sequence is shown in SEQ ID NO: 13.

Determination of the activity of SCR-3 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

(1) Construction of retrovirus vector for expression of25 mouse SCR-3

Based on the nucleotide sequence of SCR-3 ORF, SCR-3FxhoI and SCR-3Reco primers having the following

nucleotide sequences were prepared, and PCR was performed using AGM-s3-A9 cDNA as a template. An amplified fragment was inserted to the retrovirus vector pMX-IRES-GFP in the same manner as described in (1) of Example 2.

SCR-3FxhoI

ccgCTCGAGccaccATGAAGCCTTTTCATACTGCC (SEQ ID NO: 32)
SCR-3Reco

tccGAATTCttattgtttgtaggtccgtgg (SEQ ID NO: 33)

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- (2) Preparation of stromal cells highly expressing SCR-3
 AGM-s3-A7 cells in which SCR-3 was highly expressed
 were prepared by using the above retrovirus vector in
 the same manner as (2) of Example 2.
- 15 (3) Determination of activity to support hematopoietic stem cells of stromal cells in which SCR-3 is highly expressed

In the same manner as described in (III) (2) of Example 1, determination of the activity to support

20 hematopoietic stem cells was performed except that AGM-S3-A7 cells, AGM-S3-A7 cells in which SCR-3 was highly expressed through retrovirus, and AGM-S3-A7 cells into which a control vector was introduced were seeded in a 24-well culture dish (Falcon) at 1 x 10⁵ cells/well.

The results are shown in Fig. 6. Hematopoietic cells co-cultured with AGM-s3-A7 cells in which SCR-3 was highly expressed (A7/SCR-3) showed high chimerism in

recipient individuals after the transplantation compared with the parent cell lines or hematopoietic cells cocultured with the cells into which a control vector was introduced. The high chimerism was observed in myeloid and lymphoid cells two months after the transplantation. Therefore, it is revealed that hematopoietic stem cells and hematopoietic progenitor cells which can reconstitute the hematopoietic system in bodies of irradiated mice have maintained and amplified superiorly to the co-culture with cells into which SCR-3 is not 10 introduced, during the co-culture period. From the results, it is revealed that an activity of stromal cells to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor 1.5 cells is increased by high expression of SCR-3. Therefore, it is revealed that a gene product of SCR-3 has an activity to affect hematopoietic stem cells or hematopoietic progenitor cells to support survival or proliferation thereof or an activity to affect stromal 20 cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

Example 4 Cloning of SCR-4 and activity determination

25 By searching GenBank database for the nucleotide sequence shown in SEQ ID NO: 3 with BLAST, it has been found that SCR-4 has a high homology to Homo sapiens

clone 25077 mRNA of an accession number AF131820, and that SCR-4 is a mouse ortholog. This sequence is described in WO 00/66784.

The nuclotide sequence of ORF of AF131820 and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 16. Only the amino acid sequence is shown in SEQ ID NO: 17.

The nuclotide sequence of ORF of SCR-4 and the amino acid sequence deduced from the nucleotide sequence are

10 shown in SEQ ID NO: 14. Only the amino acid sequence is shown in SEQ ID NO: 15.

Determination of the activity of SCR-4 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

15 (1) Construction of retrovirus vector for expression of human SCR-4

From 3 µg of mRNA derived from fetal liver (CLONETEC, U.S.A.), cDNA was synthesized by using oligo-dT primer and reverse transcriptase (SuperscriptII, GIBCO-BRL).

- Using the cDNA as a template, the ORF region of human SCR-4 was amplified by PCR with HSCR-4FxhoI and HSCR-4RecoRV primers having the following nucleotide sequences. An amplified fragment was digested with XhoI and inserted to the retrovirus vector pMX-IRES-GFP in
- the same manner as described in (1) of Example 2. For the insertion, the pMX-IRES-GFP was digested with a restriction enzyme EcoRI, blunt-ended with KOD DNA



synthase (TOYOBO, Japan) and digested with a restriction enzyme XhoI.

HSCR-4FxhoI

CCGCTCGAGCCACCatgttggctgcaaggctggtgt (SEQ ID NO: 34)

HSCR-4RecoRV

CCGGATATCtcatttctttctgttgectcca (SEQ ID NO: 35)

- (2) Preparation of stromal cells highly expressing human SCR-4
- 10 AGM-s3-A9 cells in which human SCR-4 was highly expressed were prepared by using the above retrovirus vector in the same manner as (2) of Example 2.
 - (3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing human SCR-4, and
- 15 determination of proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay

In the same manner as described in (III) (1) 3) to

- 4) of Example 1, AGM-s3-A9 cells in which SCR-4 was
- highly expressed through retrovirus, AGM-s3-A9 cells into which a control vector was introduced, or AGM-s3-A9 cells were co-cultured with CD34-positive hematopoietic stem cells derived from human cord blood, and proliferation statuses of hematopoietic stem cells and
- 25 hematopoietic progenitor cells are determined.
 - Fig. 6 shows results when the CD34-positive hematopoietic stem cells were co-cultured with AGM-S3-A9

cells in which human SCR-4 was highly expressed, AGM-S3-A9 cells into which a control vector was introduced or AGM-S3-A9 cells for two weeks. As a result, the coculture with AGM-S3-A9 cells in which human SCR-4 was 5 highly expressed, increases of BFU-E and CFU-C were observed. Therefore, it has been revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9 increases by allowing human SCR-4 to be highly expressed. From the results, it has been revealed that human SCR-4 has 10 an activity to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor cells or an activity to affect stromal cells to impart a hematopoietic cell-supporting activity to the stromal 15 cells.

Example 5 Cloning of SCR-5 and activity determination

In the nucleotide sequence of SEQ ID NO: 4 obtained by the SBH analysis, the presence of ORF was predicted. The nuclotide sequence of ORF and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 18. Only the amino acid sequence is shown in SEQ ID NO: 19.

By searching GenBank database for the nucleotide

25 sequence of SEQ ID NO: 18 with BLAST, it has been found
that SCR-5 has a high homology with Homo sapiens
esophageal cancer related gene 4 portein (ECRG4) mRNA of

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an accession number AF325503, and that SCR-5 is a mouse ortholog of AF325503. The nuclotide sequence of ORF of AF325503 and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 20. Only the amino acid sequence is shown in SEQ ID NO: 21.

Determination of the activity of SCR-5 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

(1) Construction of retrovirus vector for expression of 10 mouse SCR-5

Based on the nucleotide sequence of SCR-5 ORF, SCR-5FxhoI and SCR-5Rblunt primers having the following nucleotide sequences were prepared for retrovirus cloning, and PCR was performed using DNA having the nucleotide sequence shown in SEQ ID NO: 23 as a template. An amplified fragment was digested with a restriction enzyme XhoI and inserted to the retrovirus vector pMX-IRES-GFP in the same manner as described in (1) of Example 2. For the insertion, the pMX-IRES-GFP was digested with a restriction enzyme EcoRI, blunt-ended with KOD DNA synthase (TOYOBO, Japan) and digested with a restriction enzyme XhoI.

SCR-5FxhoI

ccgCTCGAGccaccatgagcacctcgtctgcgcg (SEQ ID NO: 36)

25 SCR-5Rblunt

tccGTTAACttaatagtcatcatagttca (SEQ ID NO: 37)

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- (2) Preparation of stromal cells highly expressing SCR-5
 AGM-s3-A7 cells in which SCR-5 was highly expressed
 were prepared by using the above retrovirus vector in
 the same manner as (2) of Example 2.
- 5 (3) Determination of activity to support hematopoietic stem cells of stromal cells in which SCR-5 is highly expressed

In the same manner as described in (3) of Example 3, determination of the activity to support hematopoietic stem cells was performed.

The results are shown in Fig. 8. Hematopoietic cells co-cultured with AGM-s3-A7 cells in which SCR-5 was highly expressed (A7/SCR-5) showed high chimerism in recipient individuals after the transplantation compared with the parent cell lines or hematopoietic cells cocultured with the cells into which a control vector was introduced. The high chimerism was observed in myeloid and lymphoid cells two months after the transplantation. Therefore, it is revealed that hematopoietic stem cells and hematopoietic progenitor cells which can reconstitute the hematopoietic system in bodies of irradiated mice have maintained and amplified superiorly to the co-culture with cells into which SCR-5 is not introduced, during the co-culture period. From the results, it is revealed that an activity of stromal cells to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor

cells is increased by high expression of SCR-5.

Therefore, it is revealed that a gene product of SCR-5 has an activity to affect hematopoietic stem cells or hematopoietic progenitor cells to support survival or proliferation thereof or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

10 Example 6 Cloning of SCR-6 and activity determination

Based on the nucleotide sequence of SEQ ID NO: 5, a probe was prepared and AGM-s3-A9 cDNA was screened by hybridization to obtain a gene containing ORF of mouse SCR-6.

AGM-s3-A9 cells (1.4 x 10^8 cells) were dissolved in 15 20 mL of ISOGEN (Nippon gene, Japan) and total RNAs were prepared according to the attachment. Messenger RNAs were prepared from one milligram of the total RNAs according to the protocol of the mRNA purification kit 20 (Amersham Pharmacia, U.S.A.). By using SMART cDNA library construction kit (CLONTECH, U.S.A.), cDNA libraries devided to 15 fractions were prepared from the $2 \mu g$ of the prepared mRNAs according to the attachment. The libraries contained about 400,000 of independent 25 clones in total. For each fraction, PCR was performed under the following conditions to identify a fraction containing SCR-6 cDNA.

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(SEQ ID NO: 39)

Based on the sequence of a partial fragment of the mouse SCR-6 gene, the following primers were prepared, and PCR was performed with 35 cycles of 94°C, 30 seconds, 55°C, 30 seconds and 72°C, 1 minute, by using each fraction of AGM-s3-A9 cDNA libraries as a template.

SCR-6F

AGCTCATTACTGTATATTTA (SEQ ID NO: 22; 1971-1990)

(SEQ ID NO: 38)

SCR-6R

GCTATATTTCATAAGTCATC (SEQ ID NO: 22; 2330-2349)

electrophoresis and a fraction from which the PCR
product having the expected size was obtained was identified. For each of two fractions among the positive fractions, 50,000 plaques were seeded on two 15-cm petri dishes and incubated 37°C for 10 hours. Then, plaques of each petri dish were replicated to a sheet of Biodyne nylon filter (Pall, U.S.A.). The replicated nylon filter was subjected to DNA fixation treatment according to the attachment, and screening with 32P-labeled DNA probe was performed.

The probe was prepared as follows. PCR was

25 performed with 35 cycles of 94°C, 30 seconds, 55°C, 30 seconds and 72°C, 1 minute, by using SCR-6F and SCR-6R and the plasmid containing a partial fragment of the

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mouse SCR-6 gene as a template. The PCR product was subjected to 2% agarose gel electrophoresis and the amplified fragment was purified by JETSORB. By using 25 ng of the obtained PCR fragment, ³²P-labeled DNA probe was prepared with Megaprime labeling kit (Amersham Pharmacia, U.S.A.).

Hybridization and washing were performed with ExpressHybSolution (CLONETECH, U.S.A.) according to the attachment. An X-ray film was exposed to the filter and developed with a Fuji film auto developer to analyze the result. A plaque at a position corresponding to the resultant strongly exposed portion was scraped from the petri dish, and seeded again so that about 200 of plaques should appear on 10-cm petri dish. Screening was again performed according to the above-mentioned method to isolate a single plaque. The obtained clone was transfected to E. coli strain BM25.8 according to the attachment of SMART cDNA library construction kit, and the transfected cells were cultured on LB agar medium containing 50 µg/ml ampicilin to form colonies. A single colony of the transfected E. coli was inoculated to 3 ml of LB medium containing 50 µg/ml ampicilin and cultured at 30°C overnight. Plasmid was extracted with RPM kit (BIO101, U.S.A.) to obtain about 10 mg of plasmid.

Sequencing the both ends of the inserted fragment with an ABI377 DNA sequencer by using $\lambda TriplEx5'LD-$

Insert Screening Amplimer (CTCGGGAAGCGCGCCATTGTGTTGGT

(SEQ ID NO: 40); CLONTECH, U.S.A.) revealed that it
included cDNA containing the nucleotide sequence from
nucleotide 1 of SEQ ID NO: 5. The full-length

5 nucleotide sequence was also determined with the ABI377
DNA sequencer. The nuclotide sequence and the amino
acid sequence deduced from a nucleotide sequence
predicted as ORF in the nucleotide sequence are shown in
SEQ ID NO: 22. Only the amino acid sequence is shown in

Determination of the activity of SCR-6 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

(1) Construction of retrovirus vector for expression of 15 mouse SCR-6

Based on the nucleotide sequence of SCR-6 ORF, SCR-6FxhoI and SCR-6Reco primers having the following sequences were prepared for retrovirus cloning, and PCR was performed by using DNA having the nucleotide sequence shown in SEQ ID NO: 22 as a template. An amplified fragment was inserted to the retrovirus vector pMX-IRES-GFP in the same manner as described in (1) of Example 2.

SCR-6FxhoI

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25 ccgctcgagccaccATGCGTTTTTGCCTCTTCTC (SEQ ID NO: 41)

SCR-6Reco

cggaattcTTATTGGTTCACTCTGTCTG (SEQ ID NO: 42)



- (2) Preparation of stromal cells highly expressing SCR-6 AGM-s3-A9 cells in which SCR-6 was highly expressed were prepared by using the above retrovirus vector in the same manner as (2) of Example 2.
- (3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing SCR-6, and determination of proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay
- In the same manner as described in (III) (1) 3) to
 4) of Example 1, AGM-s3-A9 cells in which SCR-6 was
 highly expressed through retrovirus, AGM-s3-A9 cells
 into which a control vector was introduced, or AGM-s3-A9
 cells were co-cultured with CD34-positive hematopoietic
 stem cells derived from human cord blood, and
 proliferation statuses of hematopoietic stem cells and
 hematopoietic progenitor cells are determined.

Fig. 9 shows results when the CD34-positive hematopoietic stem cells were co-cultured with AGM-S3-A9 cells in which SCR-6 was highly expressed (A9/SCR-9), AGM-S3-A9 cells into which a control vector was introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks. As a result, the co-culture with AGM-S3-A9 cells in which SCR-6 was highly expressed, increases of BFU-E and CFU-C were observed. Therefore, it has been revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9

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increases by allowing SCR-6 to be highly expressed. From the results, it has been revealed that the gene product of SCR-6 has an activity to support survival or proliferation of hematopoietic stem cells or

hematopoietic progenitor cells or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

10 Example 7 Cloning of SCR-7 and activity determination

In the nucleotide sequence of SEQ ID NO: 6 obtained by the SBH analysis, the presence of ORF was predicted. The nuclotide sequence of ORF and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 24. Only the amino acid sequence is shown in SEQ ID NO: 25.

Determination of the activity of SCR-7 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

20 (1) Construction of retrovirus vector for expression of mouse SCR-7

Based on the nucleotide sequence of SCR-7 ORF, SCR-7FsalI and SCR-7Reco primers having the following nucleotide sequences were prepared for retrovirus cloning, and PCR was performed using DNA having the nucleotide sequence shown in SEQ ID NO: 24 as a template. An amplified fragment was inserted to the retrovirus

vector pMX-IRES-GFP in the same manner as described in (1) of Example 2.

SCR-7FSalI

acqcqtcqacccaccATGCCCCGCTACGAGTTG (SEQ ID NO: 43)

5 SCR-7Reco

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attGAATTCTCACTTCTTCCTCCTCTTTG (SEQ ID NO: 44)

- (2) Preparation of stromal cells highly expressing SCR-7 AGM-s3-A9 cells in which SCR-7 was highly expressed were prepared by using the above retrovirus vector in the same manner as (2) of Example 2.
- (3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing SCR-7, and determination of proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay
- In the same manner as described in (III) (1) 3) to 4) of Example 1, AGM-s3-A9 cells in which SCR-7 was highly expressed through retrovirus, AGM-s3-A9 cells into which a control vector was introduced, or AGM-s3-A9 cells were co-cultured with CD34-positive hematopoietic stem cells derived from human cord blood, and proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells are determined.

Fig. 10 shows results when the CD34-positive

25 hematopoietic stem cells were co-cultured with AGM-S3-A9

cells in which SCR-7 was highly expressed (A9/SCR-7),

AGM-S3-A9 cells into which a control vector was

introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks. As a result, the co-culture with AGM-S3-A9 cells in which SCR-7 was highly expressed, increases of BFU-E and CFU-C were observed. Therefore, it has been 5 revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9 increases by allowing SCR-7 to be highly expressed. From the results, it has been revealed that the gene product of SCR-7 has an activity to support survival or 10 proliferation of hematopoietic stem cells or hematopoietic progenitor cells or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

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Example 8 Cloning of SCR-8 and activity determination

By searching GenBank database for the nucleotide sequence shown in SEQ ID NO: 7 with BLAST, it has been found that SCR-8 is the same gene as Mus musculus mRNA for ADAM23 of an accession number AB009673. The nuclotide sequence of SCR-8 ORF and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 26. Only the amino acid sequence is shown in SEQ ID NO: 27.

Also, the sequence encoding Human MDC3 protein [Homo sapiens] described by JP 11155574-A has a homology of not less than 90% with SCR-8 and, therefore, is a human

ortholog of SCR-8. The nuclotide sequence of this ORF and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 28. Only the amino acid sequence is shown in SEQ ID NO: 29.

- Determination of the activity of SCR-8 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.
 - (1) Construction of retrovirus vector for expression of mouse SCR-8
- Based on the nucleotide sequence of SCR-8 ORF, SCR-8FxhoI and SCR-8Reco primers having the following nucleotide sequences were prepared, and PCR was performed using AGM-s3-A9 cDNA as a template. An amplified fragment was inserted to the retrovirus vector pMX-IRES-GFP in the same manner as described in (1) of

SCR-8FxhoI

Example 2.

ccgctcgagccaccATGAAGCCGCCCGGCAGCATC (SEQ ID NO: 45)
SCR-8Reco

- 20 cggaattcTCAGATGGGGCCTTGCTGAGT (SEQ ID NO: 46)
 - (2) Preparation of stromal cells highly expressing SCR-8 AGM-s3-A9 cells in which SCR-8 was highly expressed were prepared by using the above retrovirus vector in the same manner as (2) of Example 2.
 - (3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing SCR-8, and determination

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of proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay

In the same manner as described in (III) (1) 3) to 4) of Example 1, AGM-s3-A9 cells in which SCR-8 was highly expressed through retrovirus, AGM-s3-A9 cells into which a control vector was introduced, or AGM-s3-A9 cells were co-cultured with CD34-positive hematopoietic stem cells derived from human cord blood, and proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells are determined.

Fig. 11 shows results when the CD34-positive hematopoietic stem cells were co-cultured with AGM-S3-A9 cells in which SCR-8 was highly expressed, AGM-S3-A9 cells into which a control vector was introduced or AGM-S3-A9 cells for two weeks. As a result, the coculture with AGM-S3-A9 cells in which SCR-8 was highly expressed, increases of BFU-E and CFU-C were observed. Therefore, it has been revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9 increases by allowing SCR-8 to be highly expressed. From the results, it has been revealed that the gene product of SCR-8 has an activity to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor cells or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

CLAIMS

- 1. A DNA coding for a polypeptide of the following (A) or (B):
- (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 23 and SEQ ID NO: 25; or
 - (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
 - 2. The DNA according to claim 1, which is a DNA of the following (a) or (b):
- 15 (a) a DNA which comprises a nucleotide sequence selected from the group consisting of the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 18, the nucleotide sequence of nucleotides 630 to 1358 of SEQ ID NO: 22, and the nucleotide sequence of nucleotides 132
- 20 to 506 of SEQ ID NO: 24; or
 - (b) a DNA which is hybridizable with a DNA comprising the nucleotide sequence as defined in (a) or a prove prepared from said DNA, under the stringent condition, and which has an activity to support proliferation or
- 25 survival of hematopoietic stem cells or hematopoietic progenitor cells.
 - 3. The DNA according to claim 2, the stringent

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condition is 6 x SSC. 5 x Denhardt, 0.5% SDS and 68°C (SSC: 3 M NaCl, 0.3 M sodium citrate; 50 x Denhardt: 1% BSA. 1% polyvinyl pyrrolidone, 1% Ficoll 400), or 6 x SSC, 5 x Denhardt, 0.5% SDS, 50% formamide and 42°C.

- 5 4. A expression vector which comprises the DNA of any one of claims 1 to 3 in such a manner that the DNA can be expressed.
- 5. A cell into which the DNA of any one of claims 1 to 3 is introduced in such a manner that the DNA can be expressed.
 - 6. A polypeptide which is an expression product of the DNA of any one of claims 1 to 3, the polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
 - 7. The polypeptide according to claim 6, which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 23 and SEQ ID NO: 25, or an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence.
- 8. The polypeptide according to claim 6 or 7, which is modified with one or more modifying agents selected from the group consisting of polyethylene
 25 glycol (PEG), dextran, poly(N-vinyl-pyrrolidone), polypropylene glycol homopoymer, copolymer of polypropylene oxide/ethylene oxide, polyoxyethylated



polyol and polyvinyl alcohol.

- 9. An monoclonal antibody which binds to the polypeptide of any one of claims 6 to 8.
- 10. A method for supporting proliferation or

 5 survival of hematopoietic stem cells or hematopoietic
 progenitor cells, comprising the step of co-culturing
 stromal cells in which a DNA coding for a polypeptide of
 the following (A) or (B) is expressed, with
 hematopoietic stem cells or progenitor cells,
- 10 (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or
- 15 (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
 - 11. The method according to claim 10, wherein the DNA is a DNA of the following (a) or (b):
- (a) a DNA which comprises a nucleotide sequence selected from the group consisting of the nucleotide 25 sequence of nucleotides 1 to 1671 of SEQ ID NO: 8, the nucleotide sequence of nucleotides 1 to 1674 of SEQ ID NO: 10, the nucleotide sequence of nucleotides 1 to 366

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of SEQ ID NO: 12, the nucleotide sequence of nucleotides 84 to 1121 of SEQ ID NO: 14, the nucleotide sequence of nucleotides 1 to 1035 of SEQ ID NO: 16, the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 18, the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 20, the nucleotide sequence of nucleotides 630 to 1358 of SEQ ID NO: 22, the nucleotide sequence of nucleotides 132 to 506 of SEQ ID NO: 24, the nucleotide sequence of nucleotides 1 to 2487 of SEQ ID NO: 26, and the nucleotide sequence of nucleotides 1 to 2496 of SEQ ID NO: 28; or

- (b) a DNA which is hybridizable with a DNA comprising the nucleotide sequence as defined in (a) or a prove prepared from said DNA, under the stringent condition, and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- 12. A method for supporting proliferation or survival of hematopoietic stem cells or hematopoietic

 20 progenitor cells, comprising the step of culturing hematopoietic stem cells or progenitor cells in the presence of a polypeptide of the following (A) or (B), said polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells when the hematopoietic stem cells or hematopoietic progenitor cells are cultured in the presence of the polypeptide,

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- (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or
- (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- effect to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor

 15 cells, which comprises an effective amount of a polypeptide of the following (A) or (B), said polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells when hematopoietic stem cells or hematopoietic or hematopoietic progenitor cells are cultured in the presence of the polypeptide,
 - (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or
 - (B) a polypeptide which comprises an amino acid

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sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.

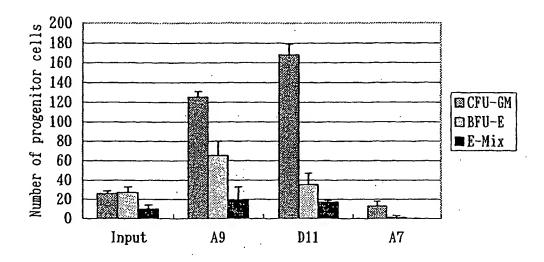


Fig.1

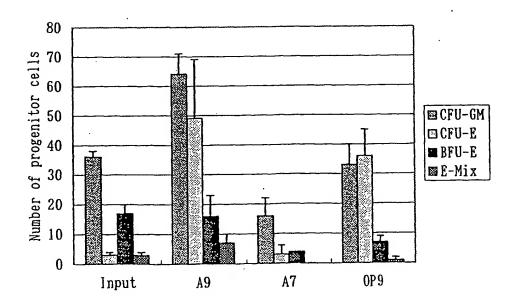
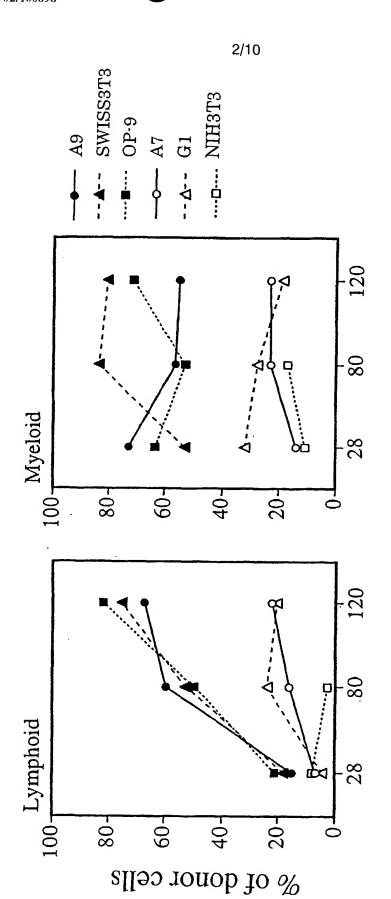
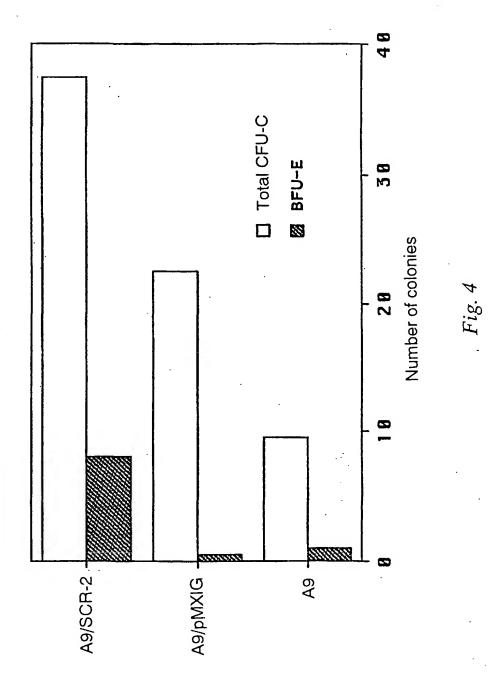
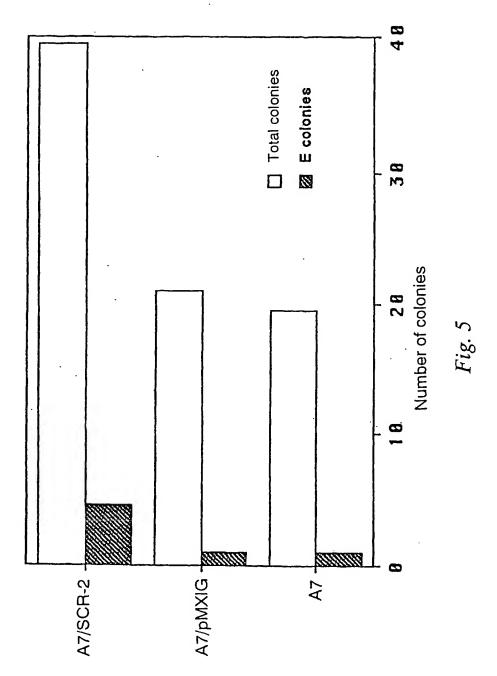


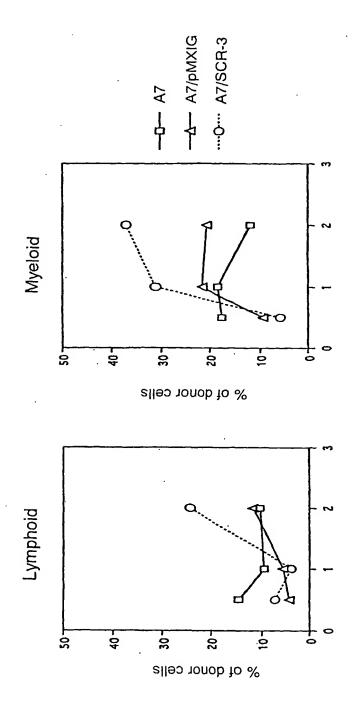
Fig.2



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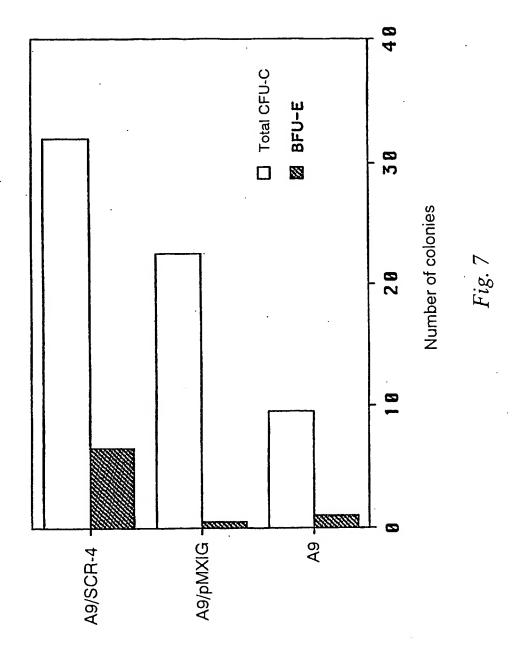


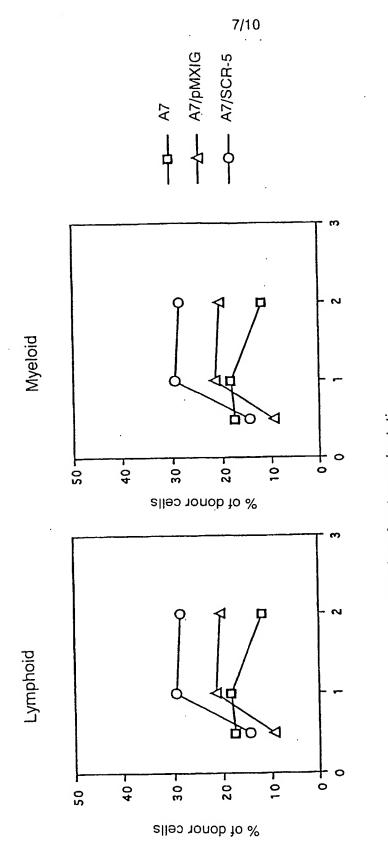




Months after transplantation

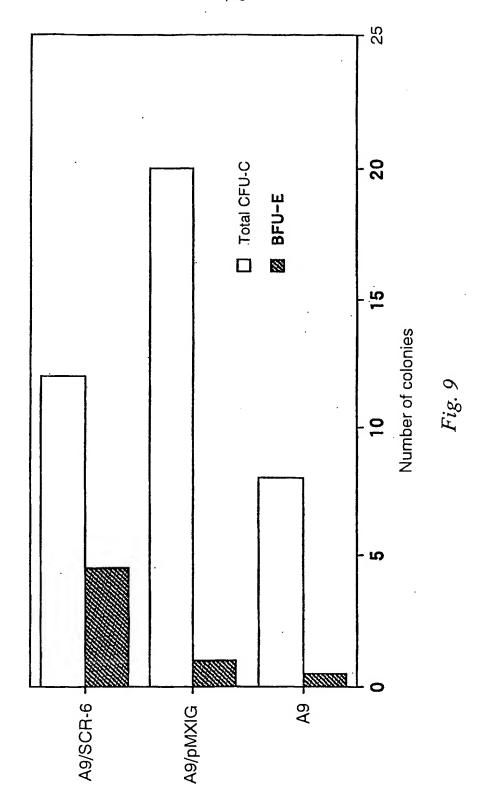
Fig 6

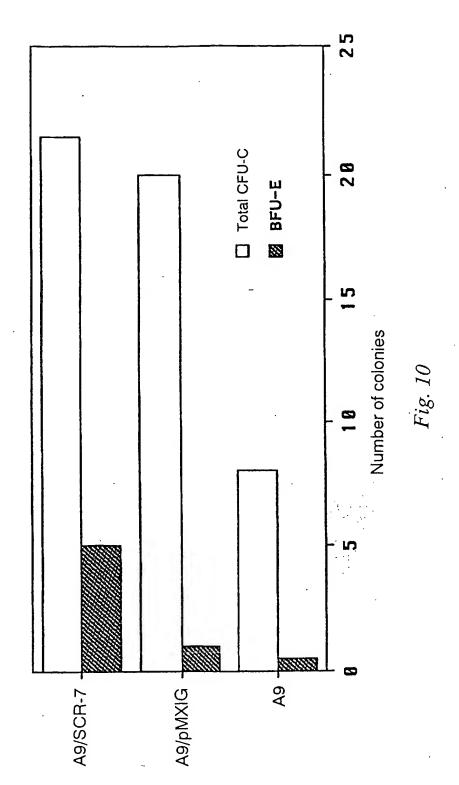


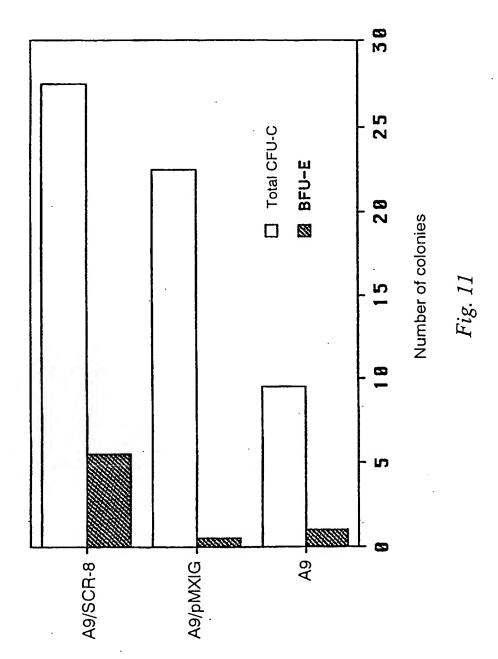


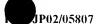
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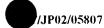
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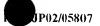
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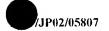
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cag gcc aca ctg gcc acc cag ctg cat ggc atc gat gac cac ttc cag Gln Ala Thr Leu Ala Thr Gln Leu His Gly Ile Asp Asp His Phe Gln	336



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gag acg ctg Glu Thr Leu	gcc gag ttc tg Ala Glu Phe Tr 165	g gca cgg ctg p Ala Arg Leu 170	ctg gag cgc c Leu Glu Arg L	tc ttc aag 528 eu Phe Lys 175
Gln Leu His	ccc cag ctg ct Pro Gln Leu Le 180		Tyr Leu Asp C	
	gag gca ctg cg Glu Ala Leu Ar			
	gcc acc cgt gc Ala Thr Arg Al 21	a Phe Val Ala		
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	gaa tgt tct cg Glu Cys Ser Ar 245			
His Cys Arg	gga gtc ccg gg Gly Val Pro Gl 260			r Cys Arg
	aaa ggc tgc ct Lys Gly Cys Le			



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								tgg Trp	Ç	960
								aca Thr 335	1(800
								ggc Gly	1()56
								gag Glu	1	104
								gcc Ala	1	152
								ctg Leu	13	200
								tgg Trp 415	1:	248
								999 Gly	13	296
					-			aag Lys	1;	344
								acc Thr	13	392
								gat Asp	14	440



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acc tgt ggc cgg agg Thr Cys Gly Arg Arg 500	gtc agc aag aag Val Ser Lys Lys 50	g agt too ago too ogg s Ser Ser Ser Ser Arg 5	Thr Pro
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tca gct gcc acc tgc Ser Ala Ala Thr Cys 530	c cca gag ccc ca s Pro Glu Pro Hi 535	c agc ttc ttc ctg cto s Ser Phe Phe Leu Leo 540	c ttc ctc 1632 J Phe Leu
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Val Pro Gln Ala Gl 50	u Ile Ser Gly G 55	lu His Leu Arg Ile C 60	ys Pro Gln
Gly Tyr Thr Cys Cy	rs Thr Ser Glu M	let Glu Glu Asn Leu A	la Asn His



65 70 75 80

Ser Arg Met Glu Leu Glu Ser Ala Leu His Asp Ser Ser Arg Ala Leu 85 90 95

Gln Ala Thr Leu Ala Thr Gln Leu His Gly Ile Asp Asp His Phe Gln 100 105 110

Arg Leu Leu Asn Asp Ser Glu Arg Thr Leu Gln Glu Ala Phe Pro Gly
115 120 125

Ala Phe Gly Asp Leu Tyr Thr Gln Asn Thr Arg Ala Phe Arg Asp Leu 130 135 140

Tyr Val Glu Leu Arg Leu Tyr Tyr Arg Gly Ala Asn Leu His Leu Glu 145 150 155 160

Glu Thr Leu Ala Glu Phe Trp Ala Arg Leu Leu Glu Arg Leu Phe Lys 165 170 175

Gln Leu His Pro Gln Leu Leu Pro Asp Asp Tyr Leu Asp Cys Leu Gly 180 185 190

Lys Gln Ala Glu Ala Leu Arg Pro Phe Gly Asp Ala Pro Arg Glu Leu 195 200 205

Arg Leu Arg Ala Thr Arg Ala Phe Val Ala Ala Arg Ser Phe Val Gln 210 215 220

Gly Leu Gly Val Ala Ser Asp Val Val Arg Lys Val Ala Gln Val Pro 225 230 235 240

Leu Ala Pro Glu Cys Ser Arg Ala Ile Met Lys Leu Val Tyr Cys Ala 245 250 255



His Cys Arg Gly Val Pro Gly Ala Arg Pro Cys Pro Asp Tyr Cys Arg 260 265 270

Asn Val Leu Lys Gly Cys Leu Ala Asn Gln Ala Asp Leu Asp Ala Glu 275 280 285

Trp Arg Asn Leu Leu Asp Ser Met Val Leu Ile Thr Asp Lys Phe Trp 290 295 300

Gly Pro Ser Gly Ala Glu Ser Val Ile Gly Gly Val His Val Trp Leu 305 310 315 320

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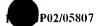
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Gly Ile Ser Lys Gly Arg Tyr Leu Pro Glu Val Met Gly Asp Gly Leu 420 425 430

Ala Asn Gln Ile Asn Asn Pro Glu Val Glu Val Asp Ile Thr Lys Pro



435

440

445

Asp Met Thr Ile Arg Gln Gln Ile Met Gln Leu Lys Ile Met Thr Asn 450 455 460

Arg Leu Arg Gly Ala Tyr Gly Gly Asn Asp Val Asp Phe Gln Asp Ala 465 470 475 480

Ser Asp Asp Gly Ser Gly Ser Gly Ser Gly Gly Gly Cys Pro Asp Asp 485 490 495

Thr Cys Gly Arg Arg Val Ser Lys Lys Ser Ser Ser Ser Arg Thr Pro 500 505 510

Leu Thr His Ala Leu Pro Gly Leu Ser Glu Gln Glu Gly Gln Lys Thr 515 520 525

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<220>

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ctg gtc gcc tgc gcc cgc ggg gac ccg gcc agc agc agg agc cgg agc tgc 96



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	ccc Pro 50															192
	tac Tyr															240
	cat His															288
	gcc Ala														-	336
	ctg Leu															384
	ttc Phe 130													-	_	432
	tca Ser															480
	acg Thr						_		_		-	_			_	528
	ctg Leu											_	_	-	-	576
	aag Lys															624



		ctg Leu														672
		ctg Leu														720
		ggc Gly									_	-	-		-	768
		tgc Cys														816
		gtg Val 275														864
		agg Arg														912
		aca Thr														960
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		gtc Val														1056
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Arg		cct Pro														1152
cag	ctc	cgc	gac	gtc	cag	gac	ttc	tgg	atc	agc	ctc	cca	999	aca	ctg	1200



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				Arg					Lys				tcc Ser 510	Arg		1536
			His					Leu					Gly		aag Lys	1584
acc Thr	tcg Ser 530	Ala	gcc	agc Ser	tgc Cys	ccc Pro 535	Gln	ccc Pro	ccg Pro	acc Thr	ttc Phe 540	Leu	ctg Leu	ccc Pro	ctc Leu	1632
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Leu Val Ala Cys Ala Arg Gly Asp Pro Ala Ser Lys Ser Arg Ser Cys 20 25 30

Gly Glu Val Arg Gln Ile Tyr Gly Ala Lys Gly Phe Ser Leu Ser Asp $35 \hspace{1cm} 40 \hspace{1cm} 45$

Val Pro Gln Ala Glu Ile Ser Gly Glu His Leu Arg Ile Cys Pro Gln 50 55 60

Gly Tyr Thr Cys Cys Thr Ser Glu Met Glu Glu Asn Leu Ala Asn Arg
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Ser His Ala Glu Leu Glu Thr Ala Leu Arg Asp Ser Ser Arg Val Leu 85 90 95

Gln Ala Met Leu Ala Thr Gln Leu Arg Ser Phe Asp Asp His Phe Gln 100 105 110

His Leu Leu Asn Asp Ser Glu Arg Thr Leu Gln Ala Thr Phe Pro Gly
115 120 125

Ala Phe Gly Glu Leu Tyr Thr Gln Asn Ala Arg Ala Phe Arg Asp Leu 130 135 140

Tyr Ser Glu Leu Arg Leu Tyr Tyr Arg Gly Ala Asn Leu His Leu Glu 145 150 155 160

Glu Thr Leu Ala Glu Phe Trp Ala Arg Leu Leu Glu Arg Leu Phe Lys 165 170 175



Gln	Leu	His	Pro 180	Gln	Leu	Leu	Leu	Pro 185	Asp	Asp	Tyr	Leu	Asp 190	Cys	Leu
Gly	Lys	Gln 195	Ala	Glu	Ala	Leu	Arg 200	Pro	Phe	Gly	Glu	Ala 205	Pro	Arg	Glu
Leu	Arg 210	Leu	Arg	Ala	Thr	Arg 215	Ala	Phe	Val	Ala	Ala 220	Arg	Ser	Phe	Val
G1n 225	Gly	Leu	Gly	Val	Ala 230	Ser	Asp	Val	Val	Arg 235	Lys	Val	Ala	Gln	Val 240
Pro	Leu	Gly	Pro	G1u 245	Cys	Ser	Arg	Ala	Val 250	Met	Lys	Leu	Val	Tyr 255	Cys
Ala	His	Cys	Leu 260	G1y	Val	Pro	Gly	Ala 265	Arg	Pro	Cys	Pro	Asp 270	Tyr	Cys
Arg	Asn	Val 275	Leu	Lys	Gly	Cys	Leu 280	Ala	Asn	Gln	Ala	Asp 285	Leu	Asp	Ala
Glu	Trp 290	Arg	Asn	Leu	leu	Asp 295	Ser	Met	Val	Leu	Ile 300	Thr	Asp	Lys	Phe
Trp 305	Gly	Thr	Ser	Gly	Val 310	Glu	Ser	Val	Ile	Gly 315	Ser	Val	His	Thr	Trp 320
Leu	Ala	Glu	Ala	Ile 325	Asn	Ala	Leu	Gln	Asp 330	Asn	Arg	Asp	Thr	Leu 335	Thr
Ala	Lys	Val	Ile 340	Gln	G1 y	Cys	Gly	Asn 345	Pro	Lys	Val	Asn	Pro 350	Gln	G1 y



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- Arg Pro Pro Ser Gly Thr Leu Glu Lys Leu Val Ser Glu Ala Lys Ala 370 375 380
- Gln Leu Arg Asp Val Gln Asp Phe Trp Ile Ser Leu Pro Gly Thr Leu 385 390 395 400
- Cys Ser Glu Lys Met Ala Leu Ser Thr Ala Ser Asp Asp Arg Cys Trp 405 410 415
- Asn Gly Met Ala Arg Gly Arg Tyr Leu Pro Glu Val Met Gly Asp Gly 420 425 430
- Leu Ala Asn Gln Ile Asn Asn Pro Glu Val Glu Val Asp Ile Thr Lys 435 440 445
- Pro Asp Met Thr Ile Arg Gln Gln Ile Met Gln Leu Lys Ile Met Thr 450 455 460
- Asn Arg Leu Arg Ser Ala Tyr Asn Gly Asn Asp Val Asp Phe Gln Asp 465 470 475 480
- Ala Ser Asp Asp Gly Ser Gly Ser Gly Ser Gly Asp Gly Cys Leu Asp
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 490
 495
- Asp Leu Cys Gly Arg Lys Val Ser Arg Lys Ser Ser Ser Ser Arg Thr 500 505 510
- Pro Leu Thr His Ala Leu Pro Gly Leu Ser Glu Gln Glu Gly Gln Lys 515 520 525
- Thr Ser Ala Ala Ser Cys Pro Gln Pro Pro Thr Phe Leu Leu Pro Leu 530 535 540



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		agt Ser 35													144
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115

120

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Met Gly Phe Gln Asp Ser Ser Asp Cys Cys Leu Ser Tyr Asn Ser Arg 50 55 60

. Ile Gln Cys Ser Arg Phe Ile Gly Tyr Phe Pro Thr Ser Gly Gly Cys 65 70 75 80

Thr Arg Pro Gly Ile Ile Phe Ile Ser Lys Arg Gly Phe Gln Val Cys 85 90 95

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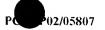
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aca cta Thr Le															161
cca ct Pro Lei												_			209
agc agg Ser Arg															257
act gga Thr Gly 60															305
atc tt: Ile Phe 75															353
gct gt Ala Va								Tyr	G1 y	Leu		Met			401
gag att Glu Ile						_	_				_		_	•	449
gat aga Asp Arg															497
aca gct Thr Ala 140	Leu												_		545



	Met															
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	ggc Gly															689
	gtt Val															737
	gca Ala 220															785
	tgt Cys															833
	ggc Gly															881
	cct Pro															929
	gga Gly															977
	atc Ile 300															1025
	ccc Pro															1073
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Phe Met Arg Val Ala Thr Met Leu Ala Thr Gly Ser Asn Arg Lys Lys 335 340 345

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<213> Mus musculus

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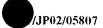
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Met Gly Arg Trp Phe Val Ala Gly Gly Ala Ala Val Gly Leu Gly Ala 85 90 95

Leu Cys Tyr Tyr Gly Leu Gly Met Ser Asn Glu Ile Gly Ala Ile Glu 100 105 110

Lys Ala Val Ile Trp Pro Gln Tyr Val Lys Asp Arg Ile His Ser Thr 115 120 125



- Tyr Met Tyr Leu Ala Gly Arg Tyr Cys Leu Thr Ala Leu Ser Ala Leu 130 135 140
- Ala Val Ala Arg Thr Pro Ala Leu Met Asn Phe Met Met Thr Gly Ser 145 150 155 160
- Trp Val Thr Ile Gly Ala Thr Phe Ala Ala Met Ile Gly Ala Gly Met 165 170 175
- Leu Val His Ser Ile Ser Tyr Glu Gln Ser Pro Gly Pro Lys His Leu 180 185 190
- Ala Trp Met Leu His Ser Gly Val Met Gly Ala Val Val Ala Pro Leu 195 200 205
- Thr Ile Leu Gly Gly Pro Leu Leu Leu Arg Ala Ala Trp Tyr Thr Ala 210 215 220
- Gly Ile Val Gly Gly Leu Ser Thr Val Ala Met Cys Ala Pro Ser Glu 225 230 235 240
- Lys Phe Leu Asn Met Gly Ala Pro Leu Gly Val Gly Leu Gly Leu Val 245 250 255
- Phe Ala Ser Ser Leu Gly Ser Met Phe Leu Pro Pro Thr Ser Val Ala 260 265 270
- Gly Ala Thr Leu Tyr Ser Val Ala Met Tyr Gly Gly Leu Val Leu Phe 275 280 285
- Ser Met Phe Leu Leu Tyr Asp Thr Gln Lys Val Ile Lys Arg Ala Glu 290 295 300
- Ile Thr Pro Met Tyr Gly Ala Gln Lys Tyr Asp Pro Ile Asn Ser Met 305 310 315 320



Leu Thr Ile Tyr Met Asp Thr Leu Asn Ile Phe Met Arg Val Ala Thr 325 330 335

Met Leu Ala Thr Gly Ser Asn Arg Lys Lys 340 345

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> 0	NA														
>	lomo	sapi	ens												
>															
> (DS														
> (1)	(103	5)												
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cac	сса	gct	ttc	acc	aag	gcc	tcc	cct	gtt	gtg	aag	aat	tcc	atc	96
		20					25					30			
220	22+	caa	taa	cto	tta	aca	cct	age	agg	ดลล	tat	acc	acc	aaa	144
Lys		UIII	ıγp	LEU	LCU	40	1.0	00,	s	0.0	45	,,,,	• • • • • • • • • • • • • • • • • • • •	2,0	
															100
aga	att	999	atc	cgg	cgt	999	aga	act	ggc	caa	gaa	ctc	aaa	gag	192
	Ile	Gly	Пe	Arg		Gly	Arg	Thr	Gly		Glu	Leu	Lys	Glu	
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															200
Alg	пр	rne		міа	шу	шу	Ala		vai	ury	LCU	019		200	
			85					30					33		
tac	tat	ggc	ttg	gga	ctg	tct	aat	gag	att	gga	gct	att	gaa	aag	336
	> 1 Control of the co	> 1038 > DNA > Homo > CDS > (1) > 16 ttg gct Leu Ala cac cca His Pro aag aat Lys Asn 35 aga att Arg Ile 50 gca ttg Ala Leu aga tgg Arg Trp tac tat	> 1038 > DNA > Homo sapi > CDS > (1) (103 > 16 ttg gct gca Leu Ala Ala cac cca gct His Pro Ala 20 aag aat caa Lys Asn Gln 35 aga att ggg Arg Ile Gly 50 gca ttg gaa Ala Leu Glu aga tgg ttt Arg Trp Phe tac tat ggc	> 1038 > DNA > Homo sapiens > CDS > (1). (1035) > 16 ttg gct gca agg Leu Ala Ala Arg	> 1038 > DNA > Homo sapiens > CDS > (1). 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Leu Glu Pro Ser Met Glu Lys Ile 70 aga tgg ttt gtt gct gga ggg gct gct Arg Trp Phe Val Ala Gly Gly Ala Ala 85 90 tac tat ggc ttg gga ctg tct aat gag	> 1038 > DNA > Homo sapiens > CDS > (1)(1035) > 16 ttg gct gca agg ctg gtg tgt ctc cgg aca Leu Ala Ala Arg Leu Val Cys Leu Arg Thr 5 10 cac cca gct ttc acc aag gcc tcc cct gtt His Pro Ala Phe Thr Lys Ala Ser Pro Val 20 25 aag aat caa tgg ctg tta aca cct agc agg Lys Asn Gln Trp Leu Leu Thr Pro Ser Arg 35 40 aga att ggg atc cgg cgt ggg aga act ggc Arg Ile Gly Ile Arg Arg Gly Arg Thr Gly 50 55 gca ttg gaa cca tcg atg gaa aaa ata ttt Ala Leu Glu Pro Ser Met Glu Lys Ile Phe 70 75 aga tgg ttt gtt gct gga ggg gct gct gtt Arg Trp Phe Val Ala Gly Gly Ala Ala Val 85 90 tac tat ggc ttg gga ctg tct aat gag att	> 1038 > DNA > Homo sapiens > CDS > (1)(1035) > 16 ttg gct gca agg ctg gtg tgt ctc cgg aca cta Leu Ala Ala Arg Leu Val Cys Leu Arg Thr Leu 5 10 cac cca gct ttc acc aag gcc tcc cct gtt gtg His Pro Ala Phe Thr Lys Ala Ser Pro Val Val 20 25 aag aat 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Thr Lys Asi 35	n Gln Trp Le	u Leu Thr Pro 40	Ser Arg G	Glu Tyr Ala ⁻ 45	Thr Lys
Thr Arg Ilo 50	e Gly Ile Ar	g Arg Gly Arg 55		Gln Glu Leu 60	Lys Glu
Ala Ala Le 65	u Glu Pro Se 70	r Met Glu Ly	s Ile Phe L 75	Lys Ile Asp	Gln Met 80
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Ile Ser Arg Thr Pro Val Leu Met Asn Phe Met Met Arg Gly Ser Trp 145 150 155 160

Val Thr Ile Gly Val Thr Phe Ala Ala Met Val Gly Ala Gly Met Leu 165 170 175

Val Arg Ser Ile Pro Tyr Asp Gln Ser Pro Gly Pro Lys His Leu Ala 180 185 190

Trp Leu Leu His Ser Gly Val Met Gly Ala Val Val Ala Pro Leu Thr 195 200 205

Ile Leu Gly Gly Pro Leu Leu Ile Arg Ala Ala Trp Tyr Thr Ala Gly 210 215 220

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48

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30/64

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Thr	Asn 50	Val	Ala	Val	Ala	G1ս 55	Asn	Thr	Ala	Lys	G1 u 60	Phe	Leu	Gly	Gly	



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45

35



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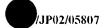
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			att Ile													701
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			ttg Leu													797
			ttc Phe 60						-			-	_			845
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			aga Arg													941
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Gly Tyr Tyr Gly His Arg Ala Pro Asp Met Asn Arg Cys Ala Arg Cys 85 90 95

Arg Ile Glu Asn Cys Asp Ser Cys Phe Ser Lys Asp Phe Cys Thr Lys 100 105 110

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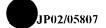
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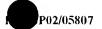
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Gly Asp Arg Trp Ile Pro Cys Ser Lys His Asp Val Phe Cys Gly Phe 645 650 655

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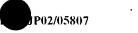
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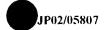
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Trp Gly Ala Ala Ala Pro Ser Ala Pro His Trp Asn Glu Thr Ala Glu 65 70 75 80

Lys Asn Leu Gly Val Leu Ala Asp Glu Asp Asn Thr Leu Gln Gln Asn 85 90 95

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Asp Thr Phe Val Tyr Met Ile Glu Pro Leu Glu Leu Val His Asp Glu 225 230 235 240

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Tyr Ser Lys Gln Met Lys Asn Leu Thr Met Glu Arg Gly Asp Gln Trp 260 265 270

Pro Phe Leu Ser Glu Leu Gln Trp Leu Lys Arg Arg Lys Arg Ala Val 275 280 285

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Ser Gly Gln Cys Pro Pro Asn Leu His Lys Gln Asp Gly Tyr Ala Cys 580 585 590

Asn Gln Asn Gln Gly Arg Cys Tyr Asn Gly Glu Cys Lys Thr Arg Asp 595 600 605

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- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW.
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Published:

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- (88) Date of publication of the international search report: 30 May 2003

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: POLYPEPTIDE HAVING AN ACTIVITY TO SUPPORT PROLIFERATION OR SURVIVAL OF HEMATOPOIETIC STEM CELL AND HEMATOPOIETIC PROGENITOR CELL, AND DNA CODING FOR THE SAME

(57) Abstract: A gene encoding a polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells is isolated by comparing expressed genes between cells which support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells and cells which do not support the proliferation or survival. Proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells is supported by using stromal cells in which the isolated gene is expressed or a gene product of the isolated gene.

A. CLASSIFICATION OF SUBJECT MATTER IPC'7. CO7K14/475 C12N15/12 CO7K14/47

C12N5/06

A61K38/00

C07K16/22

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 CO7K C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EMBL, EPO-Internal, BIOSIS, WPI Data, PAJ, SEQUENCE SEARCH, EMBASE, CHEM ABS Data

Category* Citation of document, with indication, where appropriate, of the State of	cds" 54 18 in 447 bp	1-9
9 March 2001 (2001-03-09) "Mus musculus clone MGC:7583 IMAGE:3493553, mRNA complete of Database accession no. BC00225 XP002220990 99.8% identity with SEQ ID No overlap the whole document -& DATABASE SWALL 'Online! 1 June 2001 (2001-06-01) "Hypothetical 17.0 Da protein Database accession no. Q99LS0 XP002220991	54 18 in 447 bp	
X -& DATABASE SWALL 'Online! 1 June 2001 (2001-06-01) "Hypothetical 17.0 Da proteir Database accession no. Q99LS0 XP002220991	1"	1-9
identical to SEQ ID No 19 the whole document	-/	
X Further documents are listed in the continuation of box C.	Patent family members are listed	in annex.
Special categories of cited documents: 'A' document defining the general state of the art which is not considered to be of particular relevance 'E' earlier document but published on or after the international filing date 'L' document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) 'O' document referring to an oral disclosure, use, exhibition or other means 'P' document published prior to the international filing date but later than the priority date claimed	 'T' later document published after the interpretation or priority date and not in conflict with cited to understand the principle or the invention 'X' document of particular relevance; the cannot be considered novel or cannot involve an inventive step when the document of particular relevance; the cannot be considered to involve an indocument is combined with one or ments, such combination being obvious in the art. '&' document member of the same patent 	the application but every underlying the claimed invention to econsidered to coment is taken alone claimed invention ventive step when the ore other such docupus to a person skilled family
Date of the actual completion of the international search	Date of mailing of the international se	arch report
7 February 2003	0 6. 03. 03	

Form PCT/ISA/210 (second sheet) (July 1992)

INTERNATIONAL SE H REPORT

International Application No PCT/JP 0.000

	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	Coloured to olding the
Calegory •	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DATABASE EMBL 'Online! 26 December 2000 (2000-12-26) "Homo sapiens esophageal cancer related gene 4 protein (ECRG4) mRNA, complete cds" Database accession no. AF325503 XP002220992 cited in the application 100% identity with SEQ ID No 20 in 447 bp overlap and 82.3% identity with SEQ ID No 18 in 440 bp overlap	1-9
X	the whole document -& DATABASE SWALL 'Online! 1 March 2001 (2001-03-01) "Esophageal cancer related gene 4 protein" Database accession no. Q9H1Z8 XP002220993 identical to SEQ ID No 21 and 84.5% identity with SEQ ID No in 148 aa overlap the whole document	1-9
Υ :	MOORE K A ET AL: "HEMATOPOIETIC ACTIVITY OF A STROMAL CELL TRANSMEMBRANE PROTEIN CONTAINING EPIDERMAL GROWTH FACTOR-LIKE REPEAT MOTIFS" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, NATIONAL ACADEMY OF SCIENCE. WASHINGTON, US, vol. 94, April 1997 (1997-04), pages 4011-4016, XP002915979 ISSN: 0027-8424 the whole document	1-13
Υ	WO 99 03980 A (NAKAHATA TATSUTOSHI ;KIRIN BREWERY (JP)) 28 January 1999 (1999-01-28) abstract	1-13
Y	XU M ET AL: "STIMULATION OF HUMAN PRIMITIVE HEMATOPOIESIS BY MURINE AGM-DERIVED STROMAL CELLS" BLOOD, W.B. SAUNDERS, PHILADELPHIA, VA, US, vol. 90, no. 10, 15 November 1997 (1997-11-15), page 483A XP002911189 ISSN: 0006-4971, last sentence	1-13
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C.(Continu	uation) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category *		Relevant to claim No.
Α	MOORE KATERI A ET AL: "In vitro	1-13
^	maintenance of highly purified, transplantable hematopoietic stem cells." BLOOD,	1-13
	vol. 89, no. 12, 1997, pages 4337-4347, XP002220989 ISSN: 0006-4971 the whole document	
Α	EP 0 953 354 A (FUJISAWA PHARMACEUTICAL CO) 3 November 1999 (1999-11-03) example 8	1-13
Y	DAVID G ET AL: "MOLECULAR CLONING OF A PHOSPHATIDYLINOSITOL—ANCHORED MEMBRANE HEPARAN SULFATE PROTEOGLYCAN FROM HUMAN LUNG FIBROBLASTS" JOURNAL OF CELL BIOLOGY, vol. 111, no. 6 PART 2, 1990, pages 3165-3176, XP009005399 ISSN: 0021-9525 the whole document	10-13
Υ	-& DATABASE EMBL 'Online! 4 March 1991 (1991-03-04) "Human mRNA for heparan sulfate proteaglycan (glypican)" Database accession no. X54232 XP002230116	10-13
Y	the whole document -& DATABASE SWALL 'Online! 1 February 1994 (1994-02-01) "Glypican-1 precursor" Database accession no. P35052 XP002230117 cited in the application the whole document	10-13
Υ	DATABASE EMBL 'Online! 26 September 1999 (1999-09-26) "Mus musculus glypican-1 (Gpc1) mRNA, complete cds" Database accession no. AF185613 XP002230118 cited in the application	10-13
Y	the whole document -& DATABASE SWALL 'Online! 1 May 2000 (2000-05-01) "Glypican-1" Database accession no. Q9QZF2 XP002230119 the whole document	10-13
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	INTERNATIONAL ZARCH REPORT PCT/JP 02					
C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT						
Calegory *	Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.			
Y	SCHOFIELD KAREN P ET AL: "Expression of proteoglycan core proteins in human bone marrow stroma." BIOCHEMICAL JOURNAL, vol. 343, no. 3, 1 November 1999 (1999-11-01), pages 663-668, XP002230115 ISSN: 0264-6021 the whole document ———		10-13			
			1.			

INTERNATIONAL SEARCH REPORT

International Application No PCT/JP 02/05807

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. X Claims Nos.: 2b, 11b because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically: see FURTHER INFORMATION sheet PCT/ISA/210
SEE FORTHER IN ORIGINATION SHEET FOLY 13N/210
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
see additional sheet
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
1-13 (insofar as they relate to SEQ ID Nos 8-11 and 18-21; i.e. inventions 1 and 4)
4. No required additional search tees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest. X No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet (1)) (July 1998)

INTERNATIONAL SEARCH REPORT

International Application No PCT/JP 02/05807

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

- 1. Claims: 1-13 (insofar as they relate to SEQ ID Nos 18-21) claims relating to SCR-5
- 2. Claims: 1-13 (insofar as they relate to SEQ ID Nos 22 and 23)

claims relating to SCR-6

3. Claims: 1-13 (insofar as they relate to SEQ ID Nos 24 and 25)

claims relating to SCR-7

- 4. Claims: 10-13 (insofar as they relate to SEQ ID Nos 8-11) claims relating to SCR-2
- 5. Claims: 10-13 (insofar as they relate to SEQ ID Nos 12 and 13)

claims relating to SCR-3

- 6. Claims: 10-13 (insofar as they relate to SEQ ID Nos 14-17) claims relating to SCR-4
- 7. Claims: 10-13 (insofar as they relate to SEQ ID Nos 26-29) claims relating to SCR-8

INTERNATIONAL SEARCH REPORT

International Application No PCT/JP 02/05807

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 2b, 11b

Present claims 2b and 11b relate among others to DNA which is hybridizable under stringent conditions to a probe prepared from the the nucleotide sequences of the present application. Due to the very unclear wording of the claims, such a probe can be imagined to be prepared in many ways (including possibly even nucleotide exchanges) and is thus neither defined by its length nor by its sequence. A multitude of unrelated DNA molecules can be expected to hybridize to at least one of such probe molecules.

Therefore, present claims 2b and 11b relate among others to DNA molecules only defined by reference to a desirable characteristic or property, namely the activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.

The claims cover all DNA sequences having this characteristic or property, whereas the application provides support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT for only a very limited number of such DNAs namely the sequences defined by the SEQ ID Nos themselves. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Independent of the above reasoning, the claims also lack clarity (Article 6 PCT). An attempt is made to define the DNAs by reference to a result to be achieved. Again, this lack of clarity in the present case is such as to render a meaningful search over the whole of the claimed scope impossible. Consequently, the search has not been out carried out for those parts of claims 2b and 11b which relate to DNA molecules hybridizable to probes prepared from the nucleotide sequences of the present application.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.



PCT/JP 02/05807

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-WO 9903980	Α	28-01-1999	AU WO	8243798 A 9903980 A1	10-02-1999 28-01-1999
EP 0953354	A	03-11-1999	EP US WO	0953354 A1 6495365 B1 9806422 A1	03-11-1999 17-12-2002 19-02-1998

Form PCT/ISA/210 (patent family annex) (July 1992)

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